

# DECOMP: A PDB decomposition tool on the web

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## Abstract:

The protein databank (PDB) contains high quality structural data for computational structural biology investigations. We have earlier described a fast tool (the decomp\_pdb tool) for identifying and marking missing atoms and residues in PDB files. The tool also automatically decomposes PDB entries into separate files describing ligands and polypeptide chains. Here, we describe a web interface named DECOMP for the tool. Our program correctly identifies multi-monomer ligands, and the server also offers the preprocessed ligand-protein decomposition of the complete PDB for downloading (up to size: 5GB)

**Availability:** <http://decomp.pitgroup.org>

**Keywords:** PDB, web tool, decomposition, server, ligands, SEQRES

## Background:

The Protein Data Bank [1] started to function as the depository of the crystallographic data, complementing journal publications: researchers solved the structure of a protein, wrote a paper on the result, and deposited the data of the solution in the publicly available PDB. The irregularities of the structure deposited (such as lacking atomic coordinates, broken chains, unidentified substructures) are mostly remarked in the cited publications and also in the remark-fields of the PDB file. The textual annotations in the scientific publication elsewhere or in the remark-fields in the very same PDB-file, however, make the automatic processing of the protein-structures very difficult. This statement may be a little bit confusing, since atoms, carrying the HET label are not supposed to be in the peptide-chain, so those structures that contains HET atoms other than the oxygen of the water would qualify for being a complex. Unfortunately, this is not the case. Metal ions, modified residues (in a surprisingly large number), and small molecules added in the crystallization all contain heteroatoms, and they are frequently not considered to be ligands. With our decomp\_pdb program [2] protein-ligand complexes

are identified reliably, and the ligands are deposited in separate files. Missing residues and atoms in chains are handled properly, that is, even if several atoms are missing from a chain our algorithm will still not recognize the parts as distinct chains. Placeholders are inserted into chains for missing residues/atoms (an example is given in **Figure 2**), denoting that the objects were not measured crystallographically, but - according to the more reliable sequence information - they should be there. This way our algorithm "repairs" faulty PDB's, or recognizes that flexible chain sequences are present. We should remark, that missing atoms are usually a sign of mobile loop or string in the protein-crystal, since flexible atoms will not give usable electron density maps. Consequently, mapping missing atoms this way may help to automatically identify flexible protein parts. Ligands are identified without using the HET-atom labels, properly handling modified residues and small artifacts, due to crystallization protocols. CONECT records of the ligand-atoms are computed automatically (these records for the ligands generally are not present in the PDB file).

HETATM	3303	N	GLU	G	1	15.088	10.798	23.547	1.00	14.90	N
HETATM	3304	CA	GLU	G	1	15.010	9.987	24.792	1.00	20.92	C
HETATM	3305	C	GLU	G	1	16.115	8.924	24.830	1.00	21.55	C
HETATM	3306	O	GLU	G	1	16.520	8.515	25.940	1.00	17.16	O
HETATM	3307	CB	GLU	G	1	13.635	9.327	24.908	1.00	14.23	C
HETATM	3308	CG	GLU	G	1	13.394	8.708	26.271	1.00	18.34	C
HETATM	3309	CD	GLU	G	1	12.045	8.046	26.402	1.00	18.27	C
HETATM	3310	OE1	GLU	G	1	11.293	7.936	25.435	1.00	19.98	O
HETATM	3311	OXT	GLU	G	1	16.578	8.524	23.744	1.00	21.48	O
HETATM	3312	N	BCS	G	2	11.726	7.642	27.628	1.00	23.67	N
HETATM	3313	CA	BCS	G	2	10.472	6.967	27.934	1.00	24.20	C
HETATM	3314	CB	BCS	G	2	10.726	5.484	28.206	1.00	26.79	C
HETATM	3315	SG	BCS	G	2	11.291	4.524	26.810	1.00	31.02	S
HETATM	3316	CD	BCS	G	2	9.729	3.804	26.262	1.00	32.02	C
HETATM	3317	CE	BCS	G	2	8.930	3.171	27.370	1.00	33.22	C
HETATM	3318	CZ1	BCS	G	2	7.640	3.614	27.650	1.00	35.26	C
HETATM	3319	CZ2	BCS	G	2	9.464	2.135	28.133	1.00	31.51	C
HETATM	3320	CT1	BCS	G	2	6.893	3.037	28.673	1.00	35.56	C
HETATM	3321	CT2	BCS	G	2	8.723	1.550	29.161	1.00	27.28	C
HETATM	3322	CH	BCS	G	2	7.437	2.001	29.430	1.00	30.54	C
HETATM	3323	C	BCS	G	2	9.834	7.550	29.180	1.00	22.41	C
HETATM	3324	O	BCS	G	2	10.522	8.023	30.084	1.00	21.77	O
HETATM	3325	N	PG9	G	3	8.512	7.468	29.229	1.00	21.35	N
HETATM	3326	CA	PG9	G	3	7.740	7.933	30.366	1.00	24.25	C
HETATM	3327	CB	PG9	G	3	6.555	7.062	30.633	1.00	24.94	C
HETATM	3328	CG1	PG9	G	3	5.330	7.315	30.027	1.00	25.47	C
HETATM	3329	CD1	PG9	G	3	4.250	6.459	30.220	1.00	26.21	C
HETATM	3330	CE	PG9	G	3	4.392	5.339	31.027	1.00	24.08	C
HETATM	3331	CD2	PG9	G	3	5.611	5.081	31.640	1.00	25.33	C
HETATM	3332	CG2	PG9	G	3	6.683	5.941	31.441	1.00	26.11	C
HETATM	3333	C	PG9	G	3	7.452	9.433	30.354	1.00	29.42	C
HETATM	3334	O	PG9	G	3	7.116	9.957	31.433	1.00	30.71	O
HETATM	3335	OXT	PG9	G	3	7.569	10.068	29.284	1.00	29.96	O

**Figure 1:** The DECOMP\_PDB output-ligand 10gs.pdb.out.lig.3 contains the 3-monomer GLU-BCS-PG9 molecule correctly, in one single file, even if it contains three monomer ID's.

ATOM	1636	OE1	GLN	A	209	8.145	-9.501	22.493	1.00	40.65	O
ATOM	1637	NE2	GLN	A	209	7.366	-11.004	21.011	1.00	36.70	N
ATOM	1638	OXT	GLN	A	209	5.111	-7.365	17.827	1.00	28.35	O
TER	1639		GLN	A	209						
ATOM	1640	C	MPRO	B	1M						C
ATOM	1641	CA	MPRO	B	1M						C
ATOM	1642	CB	MPRO	B	1M						C
ATOM	1643	CD	MPRO	B	1M						C
ATOM	1644	CG	MPRO	B	1M						C
ATOM	1645	N	MPRO	B	1M						N
ATOM	1646	O	MPRO	B	1M						O
ATOM	1647	N	PRO	B	2	36.456	22.522	0.112	1.00	44.99	N
ATOM	1648	CA	PRO	B	2	35.928	23.163	1.346	1.00	38.33	C
ATOM	1649	C	PRO	B	2	34.592	22.500	1.704	1.00	31.55	C

**Figure 2:** Atoms or residues are frequently missing at the beginning or at the end of polypeptide chains. In this example a missing residue and six missing atoms are identified at the beginning of chain B of pdb entry 10gs.

### Methodology:

Our program selects atoms from the PDB entry that are part of a protein or DNA chain. We do not use the chain-identifier for this purpose. However, we use SEQRES data and refined graph-theoretical algorithms described elsewhere [2]. It selects the water molecules, and removes them from the set of possible ligand atoms. Then metal and other small ions are selected, that will not be considered as ligands. A complete list of residue names that were considered as ions (so not as ligands) is given in the file `ion_list.txt`. All the remaining atoms will form the set of ligand atoms. Within this set, we use a graph-theory component detecting algorithm, so a ligand is defined as a connected component of the graph formed by the ligand atoms as vertices and the covalent bonds between the ligand atoms as the edges.

### Functionality:

The DECOMP tool correctly identifies ligand molecules, even if they are composed of more than one monomers. For example, when decomposing PDB entry 10GS with options "Export ligands", the file `10gs.pdb.out.lig.3` contains the 3-monomer GLU-BCS-PG9 molecule correctly (Figure 1).

### Utility:

Provide a list of PDB codes in the appropriate box at the web server and check the desired options. The PDB codes should be separated either by "spaces" or "new line" characters.

Press the "schedule job" button and the request will be inserted into a queue. Progress is monitored in the "Log window". The result will be a link in the "Log window" to a tar.gz file. The result file contains one directory for each of the pdb's listed. Each of these directories contains an error log with ".pdb.error" extension, the decomposed pdb file with ".pdb" extension, and if "Export ligands" or "Export ions" option was specified, then a separate file is present for each of the ligands or ions. An error file is presented if there was a fatal error while processing the PDB file. The result files are usually viewed by popular PDB viewer tools. A pre-processed, constantly updated compressed file can be downloaded with the results when the entire PDB file has been decomposed. The result files are stored for 3 days, and log files are stored for 30 days in the server.

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