Virtual screening of Indonesian herbal database as HIV-1 reverse transcriptase inhibitor

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Abstract: HIV-1 (Human immunodeficiency virus type 1) is a member of retrovirus family that could infect human and causing AIDS disease. AIDS epidemic is one of most destructive diseases in modern era. There were more than 33 million people infected by HIV until 2010. Various studies have been widely employed to design drugs that target the essential enzymes of HIV-1 that is, reverse transcriptase, protease and integrase. In this study, in silico virtual screening approach is used to find lead molecules from the library or database of natural compounds as HIV-1 reverse transcriptase inhibitor. Virtual screening against Indonesian Herbal Database using AutoDock4 performed on HIV-1 reverse transcriptase. From the virtual screening, top ten compounds were mulberrin, plucheoside A, vitexilactone, brucine N-oxide, cyanidin 3-arabinoside, alpha-mangostin, guaijaverin, erycristagallin, morusin and sanggenol N.

Keywords: Indonesian herbal database, HIV-1, molecular docking, virtual screening.

Background: HIV-1 is a member of retrovirus family that cause AIDS in human. AIDS epidemic is considered as one of the most destructive disease in the modern era. It will make immune system weakened and leading into lethal opportunistic infection [1, 2]. More than 33 million people have been infected by AIDS until 2010 and this number increases approximately by 2-3 million per year [3].

When replicating in the host cell, HIV utilizes vital production enzymes to create mature virions. One of these enzymes is reverse transcriptase (RT). HIV contains RNA genomic in its virion, and within host cell it will be converted by reverse transcriptase to cDNA (complimentary DNA) which in turn will be integrated into the host cell to produce essential protein of new viral maturation [4]. Because of RT essential role in the HIV life cycle, it has been considered as major anti HIV-1 drug targets [5]. FDA clinically approved HIV-1 reverse transcriptase inhibitors could be categorized into nucleoside reverse transcriptase inhibitors (NRTIs), nucleotide reverse transcriptase inhibitors (NtRTIs), and non-nucleoside reverse transcriptase inhibitors (NNRTIs). NRTIs were the first HIV drug approved for clinical use; it has structure similar to reverse transcriptase's substrate but lack hydroxyl functional group at position 3. In the other hand, NNRTIs target the binding site of HIV-1 reverse transcriptase directly [6].

HIV has high mutation rate as it lacks the ability to proofread its reverse transcription process. If the mutation occurred at the HIV binding pocket, it will more likely to develop resistances. Because of that reason, there is constant need for new drugs [7, 8]. Novel drugs could be developed from synthetic molecules or natural resources. Natural products have been reported as HIV-1 RT potent inhibitor, most notably from flavonoid groups [9-11]. The development of natural products compound as lead could provide benefits for HIV future treatment.

Indonesia has been considered as one of the most varied biodiversity center in the world, only outranked by Brazil in the first place. There are more than 30,000 plant species in...
This research started by selecting HIV-1 reverse transcriptase structure suitable as docking target. The HIV-1 reverse transcriptase structure consists of p66 and p51 subunit was obtained from protein databank website [18]. Inclusion criteria used to choose the target are: wild type HIV-RT, binding with ligand or inhibitor, complete chain information and has resolution less than 2.5 Å. The chosen protein structure was 3LP1, HIV-1 RT bound with NVP (nevirapine) [19]. The macromolecule downloaded as *.pdb file format.

The structure was optimized using VegaZZ [20]. The optimization includes addition of hydrogen with “protein” and “each residue ends” option, separation of solvent molecules and cofactor or ligand continued by adding partial charges (Gasteiger), applying AutoDock forcefield with AutoDockTools in MGLTools 1.5.4.

Preparing Ligand File Format
Indonesian herbal compounds structure were obtained from the Herbaldb [13]. The database 3D structure was generated using VegaZZ script “2D to 3D”. This structure then optimized by adding hydrogen with “generic organic” and “after each heavy atom” option. After adding hydrogen to the structure, the minimization was done using steepest descent and conjugate gradient 1,000 steps for each method. The last step was adding Gasteiger partial charges, and applying AutoDock forcefield with AutoDockTools.

Validating of Molecular Docking Protocol
Using 3LP1 crystal structure containing nevirapine as ligand reference, the ligand binding site coordinates were defined by AutoDockTools. This binding site will be used as the center target of molecular docking for the virtual screening. The coordinates of binding site center were x = 10.350; y = 14.076; and z = 18.252 in NNRTI pocket.

After defining the binding site, preliminary docking was done to validate the protocol using 14 control compounds given in the Table 1 (see supplementary material). The control used was referring to the FDA approved drugs. NNRTIs used as positive controls, while protease inhibitors (PIs) and integrase inhibitor as negative controls [21]. Molecular docking was attempted in triplicate using AutoDock 4.

Virtual Screening of Indonesian Herbal Database
Virtual screening was done using PyRx software with best parameters of the control compounds orientation docking. Grid parameter used was 50 x 50 x 50 units with 0.375 Å per unit. Docking parameter was set at 250,000 calculations; 27,000 generations; 150 populations; mutation and crossover rate at default. Virtual screening was applied five times.

Refinement Analysis
Top ten compounds screened were refined by afterward docking. The parameter was set at 2,500,000 calculations. Best pose then analyzed to observe its binding site and interactions using PyMOL [22]. Binding site was defined as residues in proximity of 5 Å from the docked ligand pose.

Figure 1: Best pose of top three rank from virtual screening with HIV-1 RT. Magenta: 1st rank (mulberrin), yellow: 2nd rank (plucheoside A), bluish purple: 3rd rank (vitexilactone).

Results & Discussion:
The preliminary docking was done three times (N=3) with 14 control compounds listed. Five compounds as positive controls which were known to have inhibition activity to HIV-1 RT: etravirine, nevirapine, rilpivirine, efavirenz and delavirdine. The docking results were ranked based on its best docked binding energy and shown in (Table 1). Using optimized parameter, positive controls outrank all negative controls.
After obtaining the parameter from preliminary docking, virtual screening was applied to the Indonesian Herbal Database (http://herbaldb.farmasi.ui.ac.id) [13]. Top ten compounds were given in the Table 2 (see supplementary material). The average binding energies varies from -11.28 kcal/mol to -10.36 kcal/mol. From five virtual screening runs, three of the compounds were consistently ranked as top ten from each run. The three compounds are vitexilactone, brucine N-oxide, and morusin. The first ranked compound, mulberrin, along with morusin and sanggenol N (ranked 9 and 10, respectively) were flavonoids from Moraceae family, notably from Artocarpus heterophyllus, M. orus alba, M. australis Poir, and M. mongolica [13, 23-27]. Recent research shows that flavonoids from Morus alba (e.g. morusin, kuvanon H and morusin-4'-glucoside) have anti-HIV activities [28]. However, we have not found any research that shows the anti-HIV activities of sanggenol N.

Plucheoside A ranked as second compound. It is terpene glycoside from Pluche indica [13, 23, 29]. Locher et al have studied the anti-HIV-1 activities of Hawaiian plants. From that research, it was concluded that Pluche indica could inhibit HIV-1 activities, but the mechanism of inhibition was not revealed further [30]. Compound ranked on third position is vitexilactone, a diterpene from Vitex cannabifolia, V. cannabinifolia, V. trifolia and Tinospora rumphii [13, 23, 31-33]. This plant is known well as a treatment for HIV-AIDS and shown anti HIV-1 RT activities [34]. However, there was not any recent research that directly connects vitexilactone with HIV-1 RT inhibitor activities.

Brucine N-oxide as fourth ranked compound is an indole alkaloid from Strychnos atlantica, S. lucida R. Br, S. Spinosa, and S. wallachiana [13, 23, 35]. The compound itself has no recent study which shows its activities as HIV inhibitor. Nevertheless, indole alkaloids have been researched because of its antiviral activities. Some research also show indole derived alkaloids as anti HIV, such as the study of derivatives of isatisine A done by Ji-Feng Liu et al which shown moderate anti-HIV activity [36]. The appearance of brucine N-oxide as hits could suggest us a possibility of new indole alkaloids to be anti HIV-1 RT lead compound.

Cyanidin 3-arabinoside is a flavonoid compound that spreads among plants of Ericaceae familia such as Empertrum nigrom and Vaccinium spp. Also spread amongst other plant like Theobroma cacao, M alus sylvestris, M angifera indica, and Camellia japonica [13, 23, 37, 38]. The aglycone of this compound, cyanidin and its derivatives predicted to have antitroireviral activities [39, 40].

Alpha-mangostin, ranked sixth, is a xanthone compound from Allanbackia monticola STANER L.C., Garònia cowa, G. dulcis, G. echinocarpa, G. fusca, G. mangostana, G. terpophylla, and Cratocyllum cochinhense [13, 23, 41-46]. Mangostin was known for its anti HIV activities, but the suggested inhibition mechanism was noncompetitive HIV protease inhibitor as shown in study done by Chen et al [47].

Guaijaverin is a 3-arabinopyranoside glycoside of quercetine from Psidium guajava, Taxodium distichum, and Ericaceae familia [13, 23, 48]. Eventhough it is more famous for its bacteriostatic and antiplaque activities, its aglycone was also exhibit antiviral and HIV-1 inhibitor [10, 49]. Erycristagallin is a pterocarpane from Erythrina genus plants such as Erythrina orientalis [13, 23]. The plant species of Erythrina abyssinica was suggested to have anti-HIV activities [50]. However, no recent research seems to directly relate the plant activities with erycristagallin.

Out of top ten compounds, five compounds were belong to flavonoid group or its glycoside. This could give a new view on potential development of flavonoids derived compound as anti-HIV by their inhibition mechanism of HIV-1 reverse transcriptase. A research in the future should be done to refine and enhance the better criteria for making flavonoid as HIV-1 RT inhibitor lead compound.

Mulberrin, morusin, sanggenol N, erycristagallin, and plucheoside A were amongst the top ten compounds from plants that show anti HIV-activities but not specifically show RT inhibitor activities. Further in vitro anti-RT research should be done with these compounds to find the direct connection between the plants’ compounds with their activities as anti-HIV. Anti-HIV activities of brucine N-oxide and its source plant never been reported on recent research. But as a member of indole alkaloid group, it still show some promise to be researched by in viro study as a member of indole alkaloid group. Alpha mangostin, vitexilactone, guaijaverin and cyanidin 3-arabinoside has shown anti HIV activities from previous experiment, but they could be developed further by initiating medicinal chemistry studies to improve the molecules as leads.

From further docking of the top ten compounds, we can find residues contributing as binding site and might be referred in future research. The interaction analysis of these compounds was observed on docked pose with lowest energy. All the compounds in docking process involves in hydrogen interaction with Lys101, where it serves as the hydrogen bond donor or acceptor. Some of the compounds have more than one hydrogen bond with Lys101 notably cyanidin 3-arabinoside, guaijaverin, and sanggenol N. Pose of top three best dock compound shown at (Figure 1).

Other residues that likely to have hydrogen bond with more than one top ranked compound in the best docking pose were as follow: Asn103 with vitexilactone, cyanidine 3-arabinoside, and guaijaverin; Val179 with mulberrin; Tyr181 with guaijaverin, morusin and sanggenol N; Tyr318 with mulberrin and plucheoside A; and also Glu318 from chain B with mulberrin, plucheoside A, cyanidine 3-arabinoside, and erycristagallin.

Residues that consistently within 5 angstrom boundaries from docked ligand were Leu100, Lys101, Lys102, Asn103, Val106, Val179, Ile180, Tyr181, Tyr188, Leu234, His235, Pro236, Tyr318 of p66 subunit in chain A and Glu138 p51 subunit in chain B of HIV-1 RT. This could suggest that the p51 subunit also contribute as part of the NNRTI binding site.

Conclusion:
Ten potential inhibitors against HIV-1 RT obtained from the virtual screening of Indonesian herbal database such as mulberrin, plucheoside A, vitexilactone, brucine N-oxide, cyanidin 3-arabinoside, alpha-mangostin, guaijaverin,
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erycristagallin, morusin and sanggenol N provide an opportunity for further investigation as lead compounds.

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References:
[22] The PyMOL Molecular Graphics System, Version 0.99, Schrödinger, LLC.
[38] Zapsalis C et al. J Food Sci. 1965 30: 396

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Supplementary material:

### Table 1: Docking result of control compounds using AutoDock with HIV-1 RT as target

| Rank | Name           | ΔG (N = 3) (kcal/mol) | SD (kcal/mol) | CV (%)  
|------|----------------|-----------------------|---------------|---------
| 1    | Etravirine*    | -8.82                 | 0.030         | 0.340   |
| 2    | Nevirapine*    | -7.85                 | 0.010         | 0.127   |
| 3    | Rilpivirine*   | -7.61                 | 0.221         | 2.897   |
| 4    | Efavirenz*     | -7.33                 | 0.026         | 0.361   |
| 5    | Delavirdine*   | -6.72                 | 0.516         | 7.679   |
| 6    | Raltegravir**  | -4.49                 | 2.328         | 51.887  |
| 7    | Amprenavir**   | -0.49                 | 2.829         | 573.403 |
| 8    | Tipranavir**   | 5.37                  | 8.278         | 154.146 |
| 9    | Darunavir**    | 6.55                  | 7.410         | 113.135 |
| 10   | Nelfinavir**   | 15.82                 | 14.480        | 91.528  |
| 11   | Lopinavir**    | 83.82                 | 39.737        | 47.406  |
| 12   | Saquinavir**   | 160.33                | 69.265        | 43.202  |
| 13   | Ritonavir**    | 175.55                | 82.990        | 47.273  |
| 14   | Atazanavir**   | 294.38                | 69.715        | 23.682  |

*Positive control used for this research was etravirine, nevirapine, rilpivirine, efavirenz, and delavirdine. These are members of HIV-1 NNRTIs (nonnucleoside reverse transcriptase inhibitors).

**Negative control used for this research were raltegravir (member of integrase inhibitor), amprenavir, tipranavir, darunavir, nevirapine, rilpivirine, saquinavir, ritonavir, and azatanavir (member of protease inhibitors)

### Table 2: Top ten compounds docked by AutoDock based on its binding energy with HIV-1 RT

<table>
<thead>
<tr>
<th>Rank</th>
<th>Name</th>
<th>ΔΔG (kcal/mol)</th>
<th>N</th>
<th>SD (kcal/mol)</th>
<th>CV (%)</th>
<th>Plant(s) source [13, 23]</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Mulberrin</td>
<td>-11.28</td>
<td>2</td>
<td>0.3041</td>
<td>2.697</td>
<td>Artocarpus fretessi, A. gomezianus Wallich ex Trecul, A. heterophyllus, M. alba, M. australis, M. mongolica</td>
</tr>
<tr>
<td>2</td>
<td>Plucheoside A</td>
<td>-10.82</td>
<td>4</td>
<td>0.3293</td>
<td>3.044</td>
<td>Pluchea indica</td>
</tr>
<tr>
<td>3</td>
<td>Vitexilactone</td>
<td>-10.74</td>
<td>5</td>
<td>0.0438</td>
<td>0.408</td>
<td>Vitex cannabifolia, V. cannabina, V. trifolia, Tinospora rumpiph</td>
</tr>
<tr>
<td>4</td>
<td>Brucine N-oxide</td>
<td>-10.70</td>
<td>5</td>
<td>0.0614</td>
<td>0.574</td>
<td>Strychnos atlantica, S. lucida R. Br., S. spinosa, S. wallichiana</td>
</tr>
<tr>
<td>5</td>
<td>Cyanidin 3-arabinoside</td>
<td>-10.66</td>
<td>4</td>
<td>0.1800</td>
<td>1.689</td>
<td>Mangifera indica, Acrotriche serulata, Empetrum nigrum, Epacris gunnii, Gaylussacia spp., Leucopogon collinus, Rhododendron spp., Vaccinium parvifolium, Sterculia parviflora, Theobroma cacao, Penstemon spp., Phalaris arundinacea, Polygonum spp., Aronia melanocarpa, M. sylvestris, Cinchona ledgeriana, Saxifraga spp., Camelia japonica</td>
</tr>
<tr>
<td>6</td>
<td>Alpha-mangostin</td>
<td>-10.51</td>
<td>4</td>
<td>0.2130</td>
<td>2.028</td>
<td>Allianblackia monticola STANER L.C., Garcinia kowa, G. dulcis, G. echinocarpa, G. fusca, G. Mangostana, G. terphynphyl, Cratoxylum cochinichense</td>
</tr>
<tr>
<td>7</td>
<td>Guaijaverin</td>
<td>-10.49</td>
<td>3</td>
<td>0.1270</td>
<td>1.210</td>
<td>Foeniculum vulgare, Arctostaphylos uva-ursi, Calluna vulgaris, Chamaedaphne calyculata, Richea angustifolia, R. scoparia, Hypericum erectum Thunb., Hibiscus mutabilis, Theobroma cacao L., Eucalyptus cepelocarpa, Psidium guajava, Securidaca diversifolia, Polygonium aviculare, Zanthoxylum bungeanum, Taxodium distichum</td>
</tr>
<tr>
<td>8</td>
<td>Erycristagallin</td>
<td>-10.43</td>
<td>3</td>
<td>0.3402</td>
<td>3.261</td>
<td>Erythrina abyssinica, E. crist-galli, E. orientalis, E. subumbra, E. variegata</td>
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<tr>
<td>9</td>
<td>Morusin</td>
<td>-10.43</td>
<td>5</td>
<td>0.3290</td>
<td>3.155</td>
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<tr>
<td>10</td>
<td>Sanggenol N</td>
<td>-10.36</td>
<td>4</td>
<td>0.1668</td>
<td>1.611</td>
<td>M. orus australis</td>
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