

# *In vitro* anti-acetylcholinesterase activity of an aqueous extract of *Uncaria tomentosa* and *in silico* study of its active constituents

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## Abstract:

Depletion of acetylcholine in the central nervous system (CNS) is responsible for memory loss and cognition deficit. Enzyme acetylcholinesterase (AChE) is responsible for destruction of acetylcholine (Ach) in the brain. Many herbal plant extracts have been investigated for their potential use in the treatment of Alzheimer's disease (AD) by inhibiting AChE and upregulating the levels of Ach. The current study investigated the anti-acetylcholinesterase (AChE) activity of an aqueous extract of *Uncaria tomentosa* bark which has not been reported so far in the literature. The *in vitro* study of an aqueous extract of *U. tomentosa* showed maximum inhibition of  $76.2 \pm 0.002$  % at 0.4mg/ml of final concentration with an  $IC_{50} = 0.112$  mg/ml. The mechanism of inhibition was elucidated by kinetic study which showed mixed type of inhibition, this might be due to the presence of various phytoconstituents such as oxindole alkaloids present in an aqueous extract. Based on molecular structure of phytoconstituents obtained from *U. tomentosa* known from the relevant literature, *in-silico* molecular docking study was performed against AChE protein to validate the results.

**Keywords:** Anti-cholinesterase, Acetylcholinesterase, *Uncaria tomentosa*, Ellman's assay, Enzyme Kinetics, Lineweaver-Burk plot, Oxindole alkaloids, Molecular docking, Schrodinger.

## Background:

Alzheimer's disease (AD) is a neurodegenerative disease and the most common form of dementia afflicting approximately 35 million people worldwide [1]. AD is an acquired, progressive illness with gradual deterioration of central nervous system until death. One in ten persons over 65 and nearly half of adults over 85 years have AD [2]. The clinical symptoms associated with AD include impaired ability to learn new information and recall old information, a decline in language function, dyspraxia, agnosia and impairment of executive functioning [3]. Neuropathological changes include neuronal reduction, neurofibrillary tangles, senile neurotic plaques and a variable amyloid angiopathy [4]. Neurochemical changes occur, including a marked reduction in the levels of acetylcholine and other neurotransmitters and neuromodulators [5]. Several mechanisms have been proposed to explain the cause of the disease including those of the misfolded and aggregated proteins of amyloid beta and tau [6]. But, the most conventional

theory is "cholinergic hypothesis". This hypothesis proposes that there is decline in concentration of the neurotransmitter, acetylcholine (Ach) mainly due to the action of cholinesterase enzymes in CNS [7, 8]. Therefore, the current therapeutic strategies mainly involve focusing on anti-cholinesterase inhibitors.

There are limited therapeutic options available for AD. The available drugs in the market for symptomatic treatment have several drawbacks such as side effects, low bioavailability, high cost and requirement of weekly blood monitoring [9]. In view of these limitations, the present study focuses on an aqueous extract of *Uncaria tomentosa*, commonly known as cat's claw for AChE inhibition activity. *U. tomentosa* is a large, woody vine that is indigenous to the Amazon rainforest [10]. *U. tomentosa* bark has been used traditionally in Peruvian medicine to treat gastritis, asthma and arthritis inflammatory conditions [11]. The herb has also been shown to possess antioxidant properties

[12]. *In-vivo* and *in-vitro* work has also shown that *U. tomentosa* extract alone and in combination with at least one of *Ginkgo biloba*, rosemary, gotu kola and bacopin, prevent the formation beta-amyloid plaques and the extract alone improves memory function in mice with experimental amnesia [13-14]. Of particular interest is the significant binding of an oxindole alkaloid (mitraphylline) with beta-amyloid 1-40 [15]. Number of biologically active compounds such as quinovic acid glycosides, triterpenes, flavonoids (rutin and quercetin), phytosteroids (b-sitosterol, stigmasterol, and campesterol), and catechins are present in different parts of the plant. Out of these

compounds, the most pharmacologically active compounds are tetracyclic oxindole such as rhyncophylline and isorhyncophylline and pentacyclic oxindole such as speciophylline, uncarine F, uncarine C, uncarine E, mitraphylline, isomitraphylline, pteropodine, and isopteropodine [16, 17]. The present study investigates *in-vitro* anti-AChE activity of an aqueous extract of *U. tomentosa* and *in-silico* molecular docking study of active tetra- and pentacyclic oxindole constituents, to investigate the binding interactions with acetylcholinesterase enzyme.

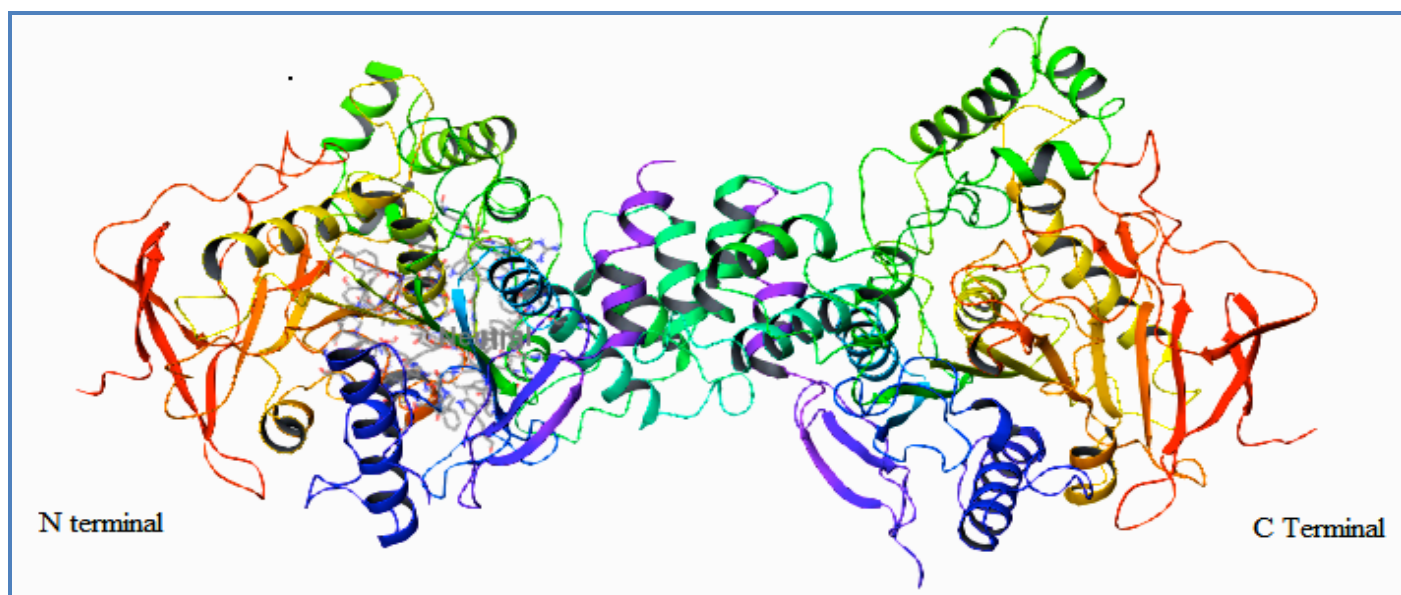


Figure 1: Three-dimensional structure of human acetyl-cholinesterase enzyme (PDBID 4EY6)

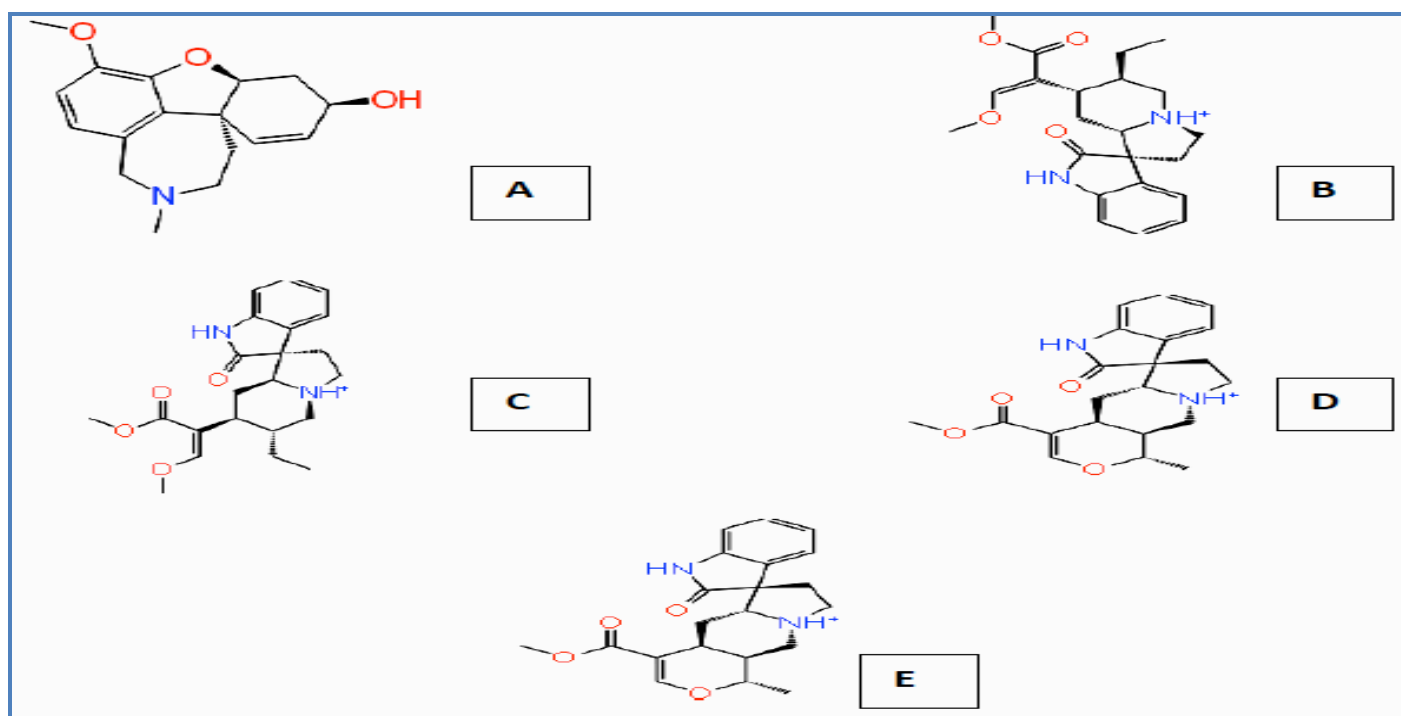


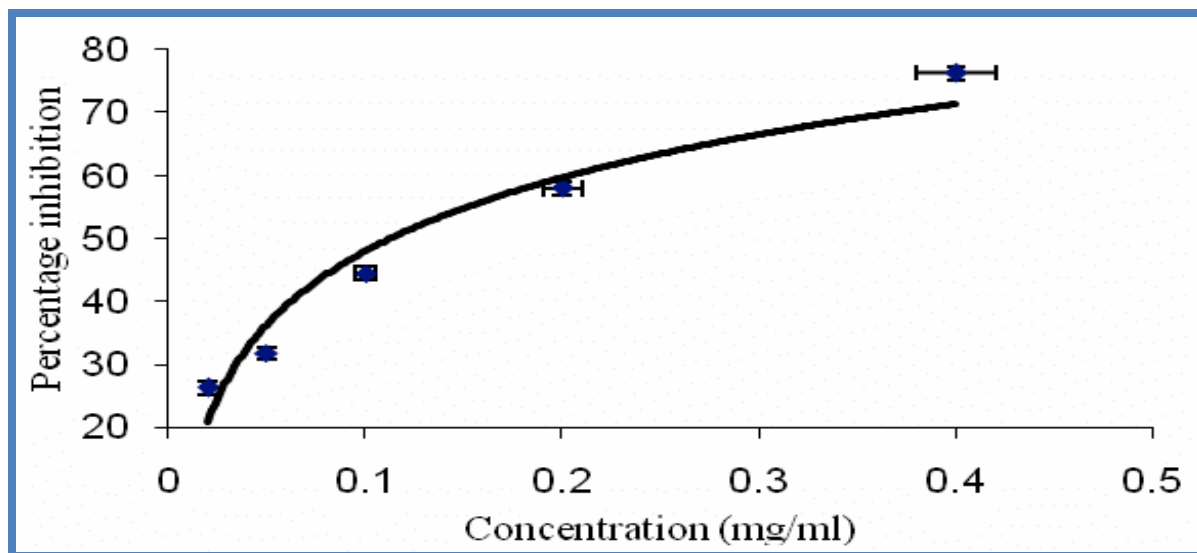
Figure 2: Two-dimension structure of A-Galantamine, B-Rhyncophylline, C-Isorhyncophylline, D-Uncarine E, E-Tacrine

**Methodology:**

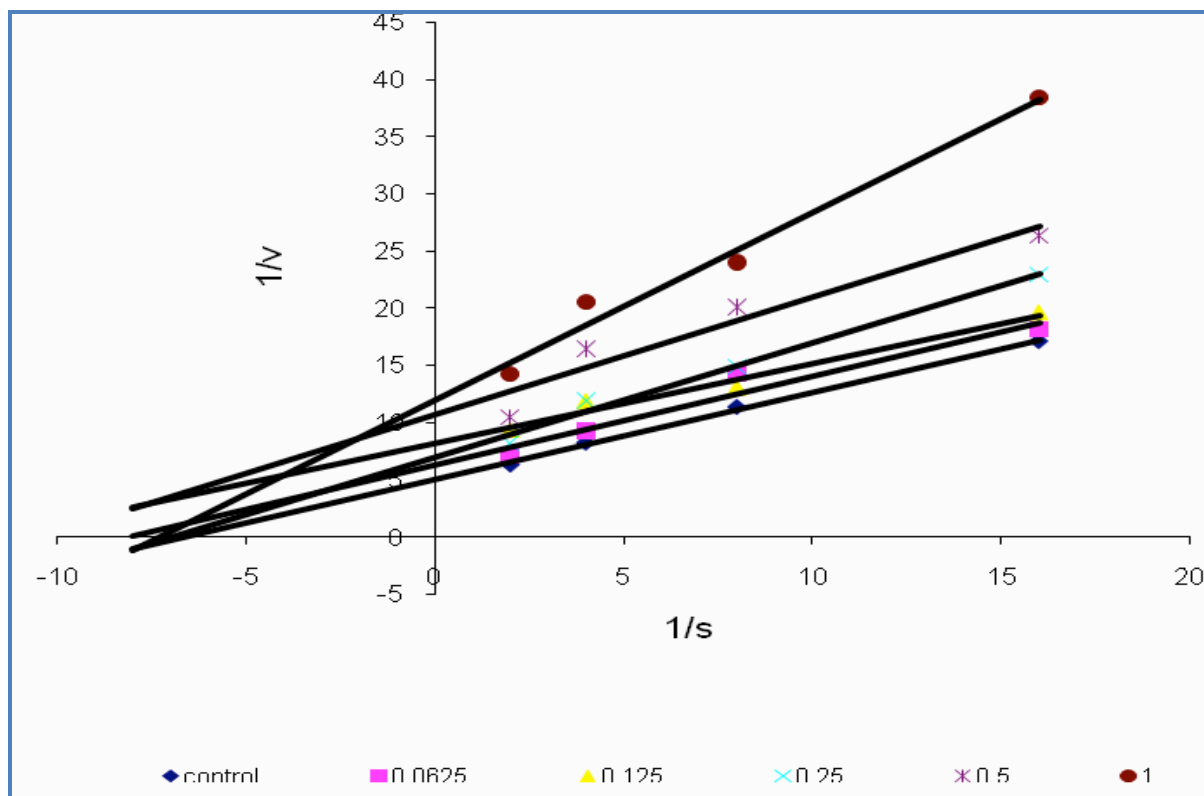
**Plant material and extraction**

The bark of *U. tomentosa* was purchased from a local store in Delhi, India and authenticated by a local botanist and a voucher specimen (USBT/SK/CC011) was stored in the herbarium at University School of Biotechnology, GGSIP University, Dwarka

Sec- 16C, New Delhi-110075. Aqueous extracts was prepared by boiling 10 gm of air dried bark powdered of herb in 50 ml (1:5 w/v) of boiling distilled water for half an hour, from which freeze dried extracts were prepared using lyophilizer (Heto, Thermo scientific) and subsequently diluted in water to the desired concentrations.



**Figure 3:** Concentration dependent enzyme inhibition activity (%) against different concentration of *U. tomentosa* aqueous extract. ( $y = 16.873\ln(x) + 86.885$ ,  $R^2 = 0.9458$ )

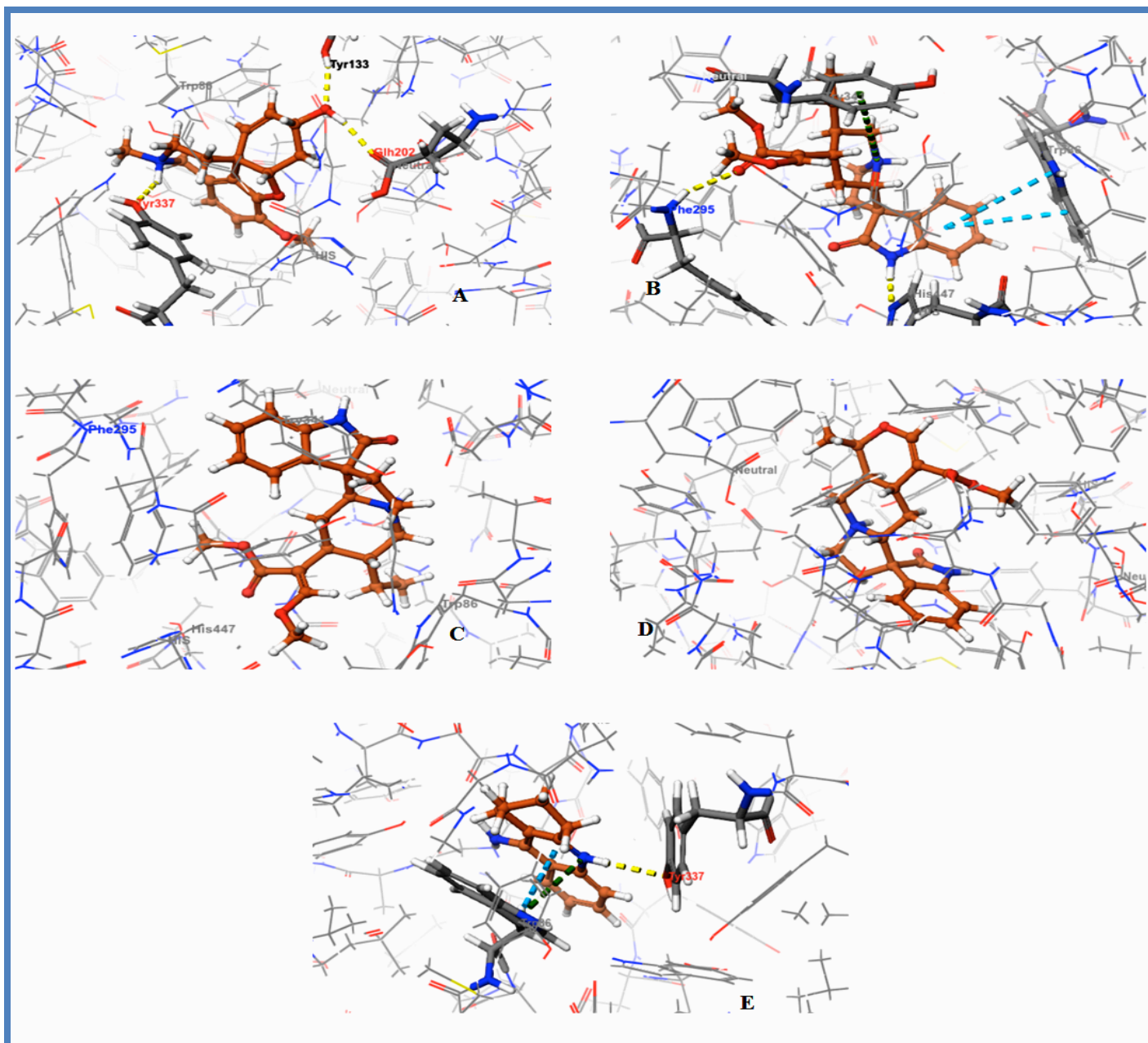


**Figure 4:** Line weaver-Burk plot of initial velocity of AChE against ATCh concentrations for control and different concentrations of *U. tomentosa*.

## In-vitro assay

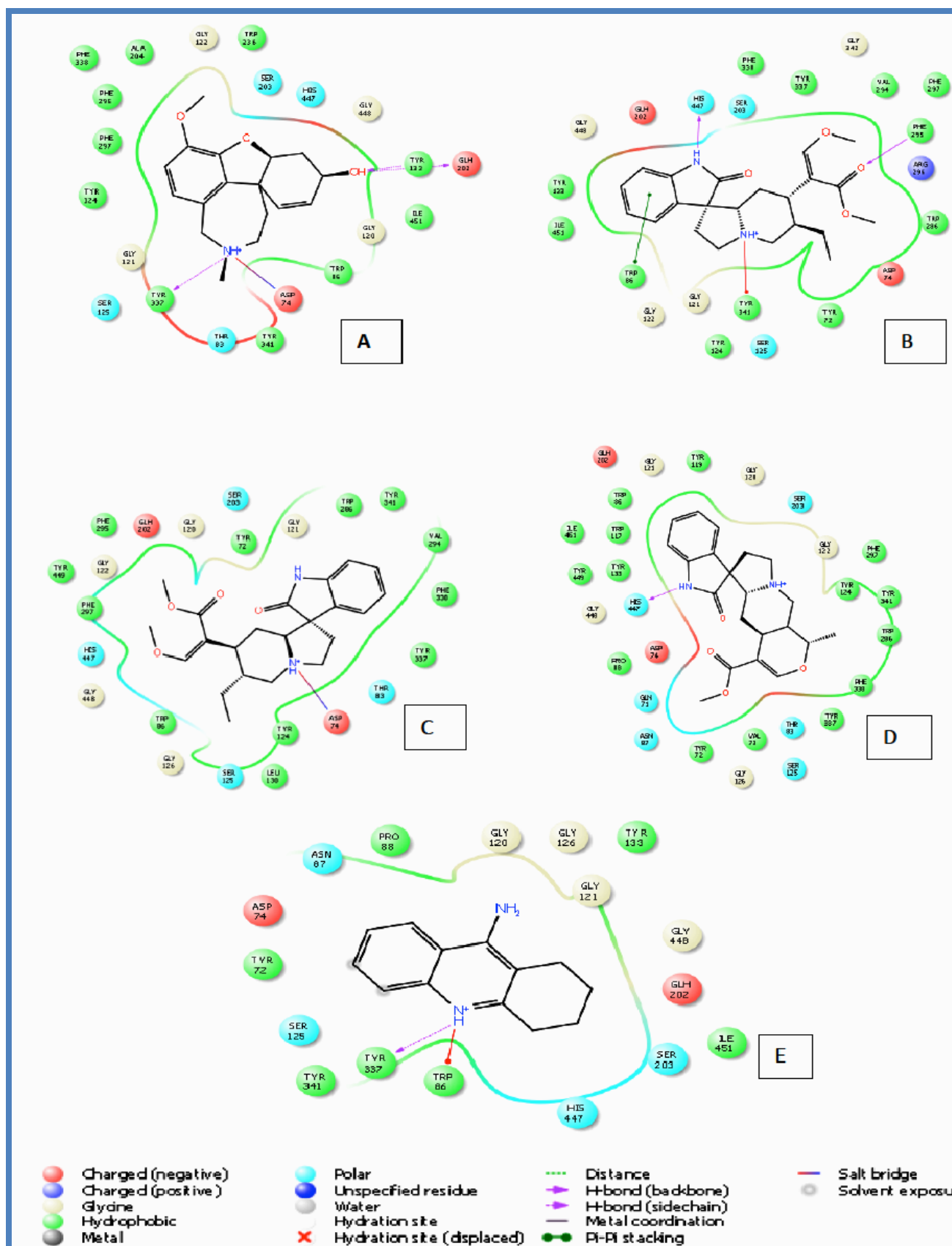
The aqueous extract of *U. tomentosa* was examined for AChE inhibitory activity using Ellman's assay [18] at various concentrations (0.4 - 0.025 mg/ml) and was dissolved in a 0.1 M phosphate buffer, pH8. To a flat bottom 96-well plate, typical run consisted of, 5  $\mu$ l of Acetylthiocholine (ATCh) (0.5 mM), 5  $\mu$ l of 5, 5'-dithiobis-2-nitrobenzoic acid (DTNB) (0.03 mM) and 5  $\mu$ l of the test extract solution at the different concentrations evaluated, which were mixed and incubated for 10 min at 30  $\mu$ C. Then, 5  $\mu$ l of AChE (0.3 U/ml) solution was added to the initial mixture to start the reaction and then absorbance was

determined at 412nm (SpectraMax M2, 96-well plate reader). A control run contained all the aforementioned constituents with exception of the test extract. All experiments were performed in triplicate with two replicates. The concentration of the tested extract that inhibited the hydrolysis of substrate ATCh by 50% ( $IC_{50}$ ) was determined by linear regression analysis. Kinetic analysis was also performed using Line weaver Burk method, where enzyme AChE was pre-incubated with different ATCh concentration ranging from 0.5-0.0625mM in the presence and absence (control) of different concentration of *U. tomentosa* ranging from 1- 0.0625 mg/ml.



**Figure 5:** Binding mode between AChE enzyme and different ligand (A-Galantamine, B-Rhyncophylline, C-Isorhyncophylline, D-Unicarine E, E-Tacrine).





**Figure 6:** Ligprot representation of interaction between different residues of AChE enzyme and ligand (A - Galantamine, B - Rhyncophylline, C - Isorhyncophylline, D - Unicarine, E - Tacrine).

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**Table 1:** Interactions in the docked complexes of ligands with AChE enzyme as obtained through Glide docking

S. No	Ligand	Glide score (Kcal/mol)	Glide Energy (Kcal/mol)	No. H-bonds	Interacting residues	Bond length (Å)
POSITIVE CONTROL						
1	Tacrine	-6.99	-29.770	1	Tyr337	2.56
2	Galantamine	-10.98	-43.850	3	Tyr133 Tyr337	2.01 2.05
PHYTOCONSTITUENTS						
1	Isorhynchophylline	-10.05	-27.436	0	- Phe295 - Trp86 ( $\pi$ - $\pi$ )	- 2.52
2	Uncarine E	-9.50	-32.934	-	-	-
3	Rhynchophylline	-9.05	-12.399	2	His447	1.92
4	Uncarine C	-7.20	-20.377		Tyr337- Tyr341 ( $\pi$ - $\pi$ )	
5	Pteropodine	-7.07	-29.155	1	His447	2.62
6	Uncarine F	-6.91	-29.116	1	His447	2.49
7	Speciophylline	-6.68	-32.934	2	Tyr337, Thr83	2.02 2.79
8	Mitraphylline	-6.00	-28.040	0	-	-
9	Isomitraphylline	-6.16	-29.606	2	Tyr337, His447 - Tyr337 ( $\pi$ - $\pi$ )	2.42
10	Isopteropodine	-5.96	-19.107	0	-	-

### Preparation of protein target structure

Acetylcholinesterase complexed with (-)-Galantamine (PDBID-4EY6, resolution 2.4 Å) was retrieved from the Protein Data Bank and further modified for Schrodinger's Glide docking calculations. For Glide v6.9 calculations, pdb file was imported to Maestro v10.4 and the protein was prepared using Protein Preparation Wizard (PPW) (Figure 1). All the water molecules were deleted except the seven water molecules, which were directly interacting with the important residues of the active site. Active site was defined using the (-)-Galantamine (inhibitor) from co-crystallized structure. The docking protocol was optimized by generating same crystal binding conformation of (-)-Galantamine with a root mean square deviation (RMSD) of 0.1273 Å [19].

### Preparation of ligands

3-D structures of *U. tomentosa* active constituents in .sdf format were retrieved from Pubchem database (Figure 2). LigPrep module of Schrodinger was used for ligand preparation [20].

### Ligand docking

Prepared protein and ligands was docked using Glide (Grid-based Ligand Docking with Energetics) v6.9 module in Schrodinger. Extra precision module of Glide algorithm was used to dock ligand to the active site. All the Glide docking runs were performed on Intel® Core™ i7-3770 CPU @ 3.40GHz of HP origin, with 4GB RAM, Windows 8Pro operating system. The output from Glide was studied in XP visualizer and images were taken to study the interaction.

### Results & Discussion:

#### Enzymatic inhibition study

An aqueous extract of *U. tomentosa* plant inhibited AChE in a concentration dependent manner (0.4 to 0.02mg/ml). The aqueous extract showed maximum inhibition of enzyme

(76.2±0.002%) at 0.4mg/ml final concentration. The IC<sub>50</sub> value obtained from inhibition curve was 0.112 mg/ml (Figure 3).

#### Kinetic study

The Lineweaver-Burk (LB) plot was used to study enzyme inhibition kinetics. The mode of inhibition displayed by LB plot was mixed inhibition (Figure 4). At the lower concentrations of an extract the plot suggested uncompetitive inhibition and at higher concentrations of extract showed non-competitive mode of inhibition. Hence, overall conclusion drawn from LB plot was mixed type of inhibition. The mixed mode of inhibition is very common in traditional medicinal plants due to the presence of different type of compounds in the extract [21]. Previous studies have suggested that for AChE induced beta-amyloid aggregation can be overcome by mixed or non competitive mode of inhibition, as these inhibitors bind to the peripheral anionic sites and therefore inhibiting beta-amyloid accumulation and aggregation [22]. Apart from antioxidant, anti-inflammatory and immunomodulatory properties *U. tomentosa* has shown to potential action on beta-amyloid plaque formation.

#### Molecular docking study

For the selected 10 phytoconstituents 27 poses were generated after ligprep, out of which the poses with better glide score and lower glide energy for each ligand was selected. Analysis of molecular docking result showed that all the phytoconstituents have their Glide score in the range of -10.5 to -5.96 Kcal/mol and Glide energy -32.934 to -12.399 Kcal/mol. Isorhynchophylline, Rhynchophylline and Uncarine E showed the best Glide score of -10.05, -9.05 and -9.50 Kcal/mol respectively. When compared with Tacrine (-6.99 Kcal/mol), all the phytoconstituents showed more and comparable Glide score except Isopteropodine, Mitraphylline and Isomitraphylline. Although, previous studies shows significant

binding of Mitrephylline to beta-amyloid (1-40) here it shows lower Glide score (-6.00 Kcal/mol). It was also observed that only Isorhynchophyllin showed Glide score as good as standard drug Galantamine, followed by Rhynchophylline. Among, the top three phytoconstituents on the basis of Glide score, Uncarine E has the lowest Glide energy (-32.934 Kcal/mol) and no hydrogen bonding interaction. While, Rhynchophylline with -12.399 Kcal/mol of Glide energy, shows two hydrogen bonding interaction with residues His447 (1.92Å bond length) and Phe295 (2.52Å) and one pi-pi interaction with Trp86. Whereas, Isorhynchophylline with Glide energy of -27.436 Kcal/mol shows no hydrogen bonding interaction (**Figures 5 & 6**). To sum up all the phytoconstituents of *U. tomentosa* more or less shows a good binding affinity towards the active site of the acetylcholinesterase, on the basis of Glide score and Glide energy (**Table1**).

### Conclusion:

*U. tomentosa* is an interesting herb in terms of its potential use in the treatment of AD. Previous *in-vitro* study showed that *U. tomentosa* extract prevents beta-amyloid plaque formation and in *in-vivo* improves memory function in mice. Our study revealed that an aqueous extract of *U. tomentosa* exhibits potent anti-AChE activity might be due to the presence of active constituent's oxindol alkaloids. Kinetic studies have indicated that an aqueous extract showed mixed mode of inhibition due to the presence of other phytoconstituents, which need further analysis. Above finding was validated with molecular docking study of its active constituents. Out of all the alkaloids Isorhynchophylline, Rhynchophylline and Uncarine E showed highest Glide score and lowest Glide energy as good as that of Galantamine the standard drug approved by food and Drug Administration (FDA). Moreover, *U. tomentosa* antioxidant, antiinflammatory, immuno-modulatory, anti AChE and potential amyloid plaque prevention properties may have combined benefits for the treatment of AD. These findings add to a body of evidence suggesting further evaluation of the effect of aqueous extracts of *U. tomentosa* and its constituents.

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