

Structural modelling and phylogenetic analyses of *PgeIF4A2* (Eukaryotic translation initiation factor) from *Pennisetum glaucum* reveal signature motifs with a role in stress tolerance and development

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Abstract:

Eukaryotic translation initiation factor 4A (*eIF4A*) is an indispensable component of the translation machinery and also play a role in developmental processes and stress alleviation in plants and animals. Different *eIF4A* isoforms are present in the cytosol of the cell, namely, *eIF4A1*, *eIF4A2*, and *eIF4A3* and their expression is tightly regulated in cap-dependent translation. We revealed the structural model of *PgeIF4A2* protein using the crystal structure of *Homo sapiens eIF4A3* (PDB ID: 2J0S) as template by Modeller 9.12. The resultant *PgeIF4A2* model structure was refined by PROCHECK, ProSA, Verify3D and RMSD that showed the model structure is reliable with 77 % amino acid sequence identity with template. Investigation revealed two conserved signatures for ATP-dependent RNA Helicase DEAD-box conserved site (VLDEADEML) and RNA helicase DEAD-box type, Q-motif in sheet-turn-helix and α -helical region respectively. All these conserved motifs are responsible for response during developmental stages and stress tolerance in plants.

Keywords: *eIF4A2*, Eukaryotic initiation factor, helicase, structural modelling, *Pennisetum glaucum*

Background:

Pennisetum glaucum (L.) R. Br.) has not been extensively utilized for its genes providing abiotic stress tolerance including eukaryotic translation initiation factors (eIFs) that are involved in the alleviation of salinity and other environmental stresses in plants and animals. *eIF4A2* is a member of the family of DEAD-box RNA helicase which is the largest family of RNA helicases. The number of these proteins in *Pennisetum glaucum* is unknown. A soybean DEAD-box RNA helicase was found to be induced in presence of salinity stress [1]. A LOS4 RNA helicase was found to have activity in cold stress response of *Arabidopsis* [2].

Presence of at least a single isomer of *eIF4A* is mandatory for proper translation and mediation of response during abiotic and biotic stresses. An insight into the structure of *PgeIF4A2* would give us a fair idea of its elements responsible for its role in

various developmental and stress responses. Here, we use structural modelling technique that is widely employed, and aims at predicting the 3D structure of bio-molecules, relying largely on resources such as pattern/function and sequence. There is very few data regarding the structural modelling of RNA helicases in plants that are responsible for enhanced tolerance to abiotic stress. Knowledge of the-(3D) structure of *eIF4A2* from *Pennisetum glaucum* could pave way for better engineered crops tolerant to abiotic stress. However, 3D structure of *eIF4A2* from *Pennisetum glaucum* is unknown till date. In the present study, we generate the 3D structure of *eIF4A2* from *Pennisetum glaucum* based on the available human *eIF4A3* (2J0S) structural homologue as template. The model structure *PgeIF4A2* was validated with standard parameters (PROCHECK, PROSAIL, Verify3D, RMSD). This study could prove useful in functional characterization of plant RNA helicases in response to plant

stress tolerance. These particular domain sequences could be used with newer technologies like CRISPR to determine their roles in mediating both abiotic and biotic stress response mechanisms in plants.

Methodology:

Sequence Analysis:

Our lab has isolated the nucleotide sequence for *PgeIF4A2* from *Pennisetum glaucum*. It was found to be over-expressed in a cDNA library under salinity stress and it was submitted to NCBI under the Accession Number EU856535.1 [3]. Structural modelling is initiated by looking in the Protein Data Bank for already known protein structures and the target sequence is used as the query [4]. In this search, the *PgeIF4A2* is compared with the sequence of each of the structures present in the database. Using PDB BLAST, the *PgeIF4A2* was searched for a similar sequence [5] against the PDB. The BLAST results yielded X-ray structure of 2J0S, the *eIF4A* from *Homo sapiens* with 77 % similarity to *PgeIF4A2*.

Modelling:

Modeller 9.12 was used to generate the theoretical structure of *PgeIF4A2* from *P. glaucum*. Protein structure prediction was done by comparative modelling. Various possible kinds of information about the target sequence were used by this program.

1	MAAXITSRRGAGASRSMDDENLIT---FETSPGVEVWSSFDQMGIKDDLRL	47
1	ATTATMAISGASARKRLKKEEDMTKVEFETSEEVDVPTPTFDITMGLREDDLRL	50
48	GIYGYGFEKPSAIQQRAVLPPIINGRDVIAQAQSGTGKSSMISLTVQIVD	97
51	GIYAYGFEKPSAIQQRAIKQIIKGRDVIQAQSQSGTGKATATFSISVQLCLD	100
98	TAVRVEQALILSPTRELASQTEKVMVAVGDYLNQVHACIGGKKSISEDIR	147
101	IQVRETQALILAFPTRELAVQIQKGLLALGDVYMNVCCHACIGGTINVEDIR	150
148	RLENGVHVVSPTGPRVCDMIKRRILRTRAIKLI	197
151	KLDYGGQHVVAGTPGRVDFMIRRRSLRTRAIKMI	200
198	YDVYRYLPPPELQVLLISATLPHEILEMISKEMTEFVRIILVKRDELTLLEGI	247
201	YDVYRYLPPPATQVLLISATLPHEILEMTNKFMTDFIRILVKRDELTLLEGI	250
248	KQFFVAVEKEEWFDTLCLDLYDILTITQAVIFCNKIKRQVWLTERRMSNN	297
251	KQFFVAVEREEWKFDTLCLDLYDILTITQAVIFCNKIKRQVWLTERRMREAN	300
298	FTVSAHMGDMPQQRDAIMTEFRSGATRVLITTDVWARGLDVQVSLVIN	347
301	FTVSSMHGDMPPQKERESIMKEFRSGASRVLITTDVWARGLDVPPVSLIIN	350
348	YDLFNNRELYIHRIGRSGRFGKGVAINFVFKDDIRILRDIEQYYSTQID	397
351	YDLFNNRELYIHRIGRSGRYGRKGVAINFVKNDDIRILRDIEQYYSTQID	400
398	EMPMNVADLI	407
401	EMPMNVADLI	410

Figure 1: Alignment of deduced amino acid sequences of eukaryotic translation initiation factors from *Homo sapiens* (*H. sapiens*; template) and *Pennisetum glaucum* (*P. glaucum*; target) revealing the signature motifs for ATP-dependent RNA helicase and RNA helicase Q-motif binding employing pair-wise alignment tool.

Model validation of *PgeIF4A2*

Model evaluation:

PROCHECK, ProSA-Web [6] and Verify 3d [7] were used to evaluate the model on the basis of geometrical and stereo-chemical constraints.

Root Mean Squared Deviation:

In general terms, RMSD is used to represent the distance between two objects. In case of structural determination, this value is indicative of the degree of similarity between two three dimensional structures. Maximum similarity of structures is represented by a lower value. The RMSD value between the template 2J0S and *PgeIF4A2* was calculated using MOE.

Phylogenetic analysis:

Molecular Evolutionary Genetic Analysis (MEGA) software (version 4.0.02) [8], was used for carrying out the phylogenetic analysis of the sequences using N-J method with used bootstrap value 5,000 replicates.

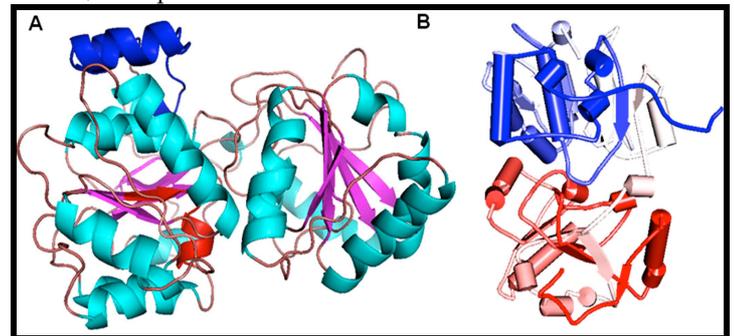


Figure 2: (A) Structural modelling of *PgeIF4A2* showing ATP-dependent RNA helicase DEAD-box conserved domain and RNA helicase DEAD-box type, Q-motif in red and blue respectively as visualized by Pymol. (B) Cartoon representation of *PgeIF4A2* model showing its N- and C- termini in red and blue, respectively as visualized by Discovery Studio.

Result & Discussion:

Modelling of *PgeIF4A2*:

The *PgeIF4A2* protein sequence comprises of 407 amino acid residues. The structural searching using PDB blast analysis revealed 77% sequence identity to *PgeIF4A3* from *H. sapiens* (PDB ID : 2J0S) with an e-value of 0.0. ScanProsite server identified the fragment VLDEADEML as the ATP-dependent RNA Helicase DEAD-Box conserved site signature motif (181-189 residues) as a consensus pattern in both. Sequence alignment revealed that RNA helicase DEAD-box type, Q-motif was also 80% conserved in both target and template *eIF4A* sequences (Fig. 1 & 2). Further, we developed three model structures for *PgeIF4A2* using Modeller 9.12. Further refinement and validation was done with the model with the lowest Discrete Optimized Protein Energy, a statistical potential used to assess structural models) score of -324317.04 as it was considered to be thermodynamically stable. It was latter visualized by PyMOL Molecular Graphics System.

Validation of *PgeIF4A2* structure:

Modeller 9.12 was used for evaluation of the stereo-chemical quality and accuracy of the predicted model and this is based on Ramachandran plot calculation. Good stereo-chemical quality of the models is generally represented by a score which is close to 100 %. Torsion angles ϕ and ψ were used to examine the reliability of the *PgeIF4A2*. The protein structure was evaluated

by a percentage quality measurement which included four kinds of occupancies: core, allowed, generously allowed and disallowed regions. The Modeller 9.12 generated model revealed 88.4 % residues falling in favoured region, 10.6 % residues in allowed region, and 1.0 % residues in outlier region while no residues were found in the disallowed region of the Ramachandran plot.

ProSA-Web analysis of the model gives a Z-score value of the *PgeIF4A2*. The Z-score is indicative of the overall quality of *PgeIF4A2* protein structure. Using it, the deviation of the total energy of the *PgeIF4A2* is measured with respect to an energy distribution derived from random conformations. For a clear interpretation of the Z-score of the specified protein, its particular value is displayed in a plot which also contains the Z-score of all experimentally determined protein chains in *PgeIF4A2*. Different colors (NMR in dark blue and X ray in light blue) are used to differentiate groups of structures from different sources (X-ray, NMR). Another application of this plot is to ascertain whether the Z-score of the *PgeIF4A2* is within the range of scores typically found for proteins of similar size belonging to one of these groups. Z-score value (-9.66) of the *PgeIF4A2* is located within the space of proteins determined by X ray crystallography and NMR. This value was extremely close to the value of template *HseIF4A3* (-9.57), which inferred that the model thus obtained was reliable and very close to experimentally determined structures. Verify3D showed 94.87 % of the residues had a score greater than 0.2 that corresponded for a good quality model. RMSD was performed between equivalent atom pairs to measure the degree of structural similarity. RMSD analysis of the *PgeIF4A2* model was measured from its *HseIF4A3* (2J05) from *Homo sapiens* using MOE software. The Ca RMSD and backbone RMSD deviation for the *PgeIF4A2* model and the *HseIF4A3* template crystal structure were 1.02 Å, and 1.03 Å respectively and over all RMSD was 1.61 Å (Fig. 3). Thus, the *PgeIF4A2* model generated by Modeller 9.12 was confirmed to be reliable and accurate.

Comparative structural alignment of *PgeIF4A2*

Alignment of *PgeIF4A2* was done with its homologous sequences and it revealed that 15 helices and 14 strands were conserved in all the selected species (Fig. 4). High sequence similarity was observed in the entire sequence. The ATP-dependent RNA Helicase DEAD-box conserved site and RNA helicase DEAD-box type, Q-motif signature motifs were present in sheet-turn-helix and right handed alpha helix regions respectively (Fig. 3). Our analysis revealed that the alpha helix might be involved in helicase activity. In addition, it revealed that the ATP-dependent RNA Helicase DEAD-box signature consensus pattern VLDEADEML) was conserved among all sequences. Thus, the model provides insight into the molecular function of the two conserved helicase domains in *PgeIF4A2* in response to stress.

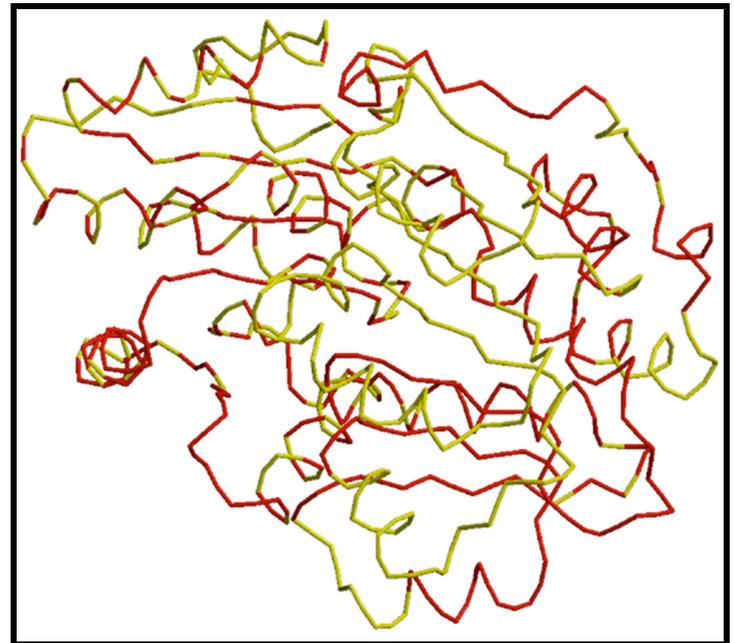


Figure 3: Superposition of Ca backbone of *PgeIF4A2* (target) and *HseIF4A* (template) by SuperPose Version 1.0 and visualized by Pymol.

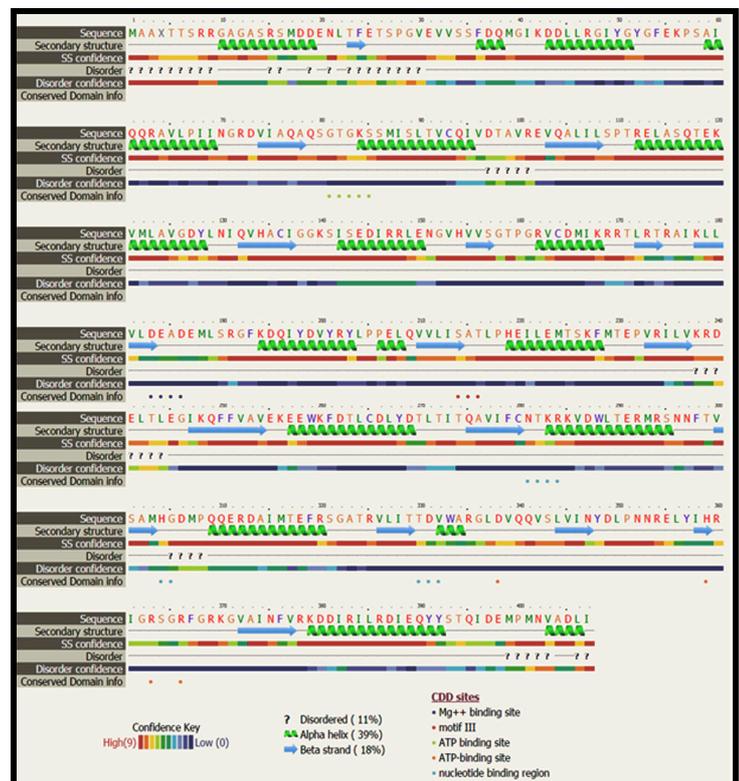


Figure 4: Comparative structure sequence alignment of deduced amino acid sequences of *PgeIF4A2* along with other homologues.

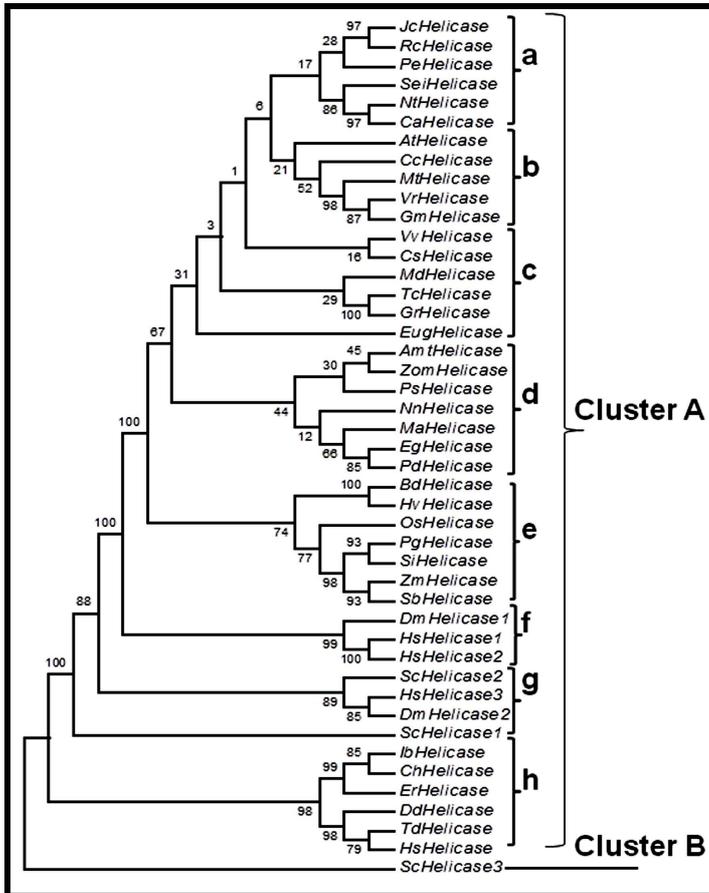


Figure 5: Phylogenetic analysis showing similarity of *Pennisetum glaucum* (*PgeIF4A2*) with other *eIF4As* from plants (dicots and monocots), mammals and microbes employing Molecular Evolutionary Genetics Analysis (MEGA) software version 4.0.

Phylogenetic analysis:

The RNA helicase family *PgeIF4A2* is highly conserved among all higher plants, animals and prokaryotic organisms. It is a multi-gene family with each gene having multiple isoforms. Duplication and divergence led to the ubiquity of the *PgeIF4A2* helicase family. To decipher the phylogenetic relationship of *PgeIF4A2* with other plant, mammal and microbial orthologs, protein sequences available in the NCBI database were selected. *eIF4As* from seven monocots, twenty four dicots, and fourteen microbes and mammals were chosen for the phylogenetic analysis. The analysis (Fig. 5) showed a clear demarcation of these *eIF4As* into two prominent clusters: cluster A and B. Cluster

B comprised of only *Saccharomyces cerevisiae* and the rest were grouped in cluster A. Further, cluster A was heavily divided into sub groups. The subgroup a, b, c and d of cluster A included all the dicots while subgroup e had all the monocots including our target *Pennisetum glaucum eIF4A*. Subgroup f, g and h included all the microbial and mammal *eIF4As*. Therefore, our target *eIF4A* is closer phylogenetically to the subgroup including human helicases 1 and 2. However, *PgeIF4A2* is closest to *Oryza sativa* and *Setaria italica* helicases. All the major clusters gave bootstrap values higher than 60. The tree showed distinct crop-specific clustering of sequences, revealing clear crop specific sequence differences.

Conclusion:

Eukaryotic translation initiation factors are a key component for the regulation of translational machinery and also play a significant role in providing stress tolerance to plants. Therefore, the model structure of *PgeIF4A2* will be extremely crucial in establishing its molecular function in response to plant stress. To the best of our knowledge this is the first report regarding the structural modelling of *Pennisetum glaucum PgeIF4A2*. This model provides a perception into the molecular functions of ATP-dependent RNA Helicase DEAD-box conserved site, helicase C-terminal and RNA helicase Q motif signature motifs during stress resistance in plants.

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