



Protein targets in red complex pathogens for catechin

T. Cibikkarthik, A.S. Smiline Girija & J. Vijayashree Priyadharsini*

Saveetha Dental College, Saveetha Institute of Medical and Technical Sciences (SIMATS), Saveetha University, Chennai, India; J. Vijayashree Priyadharsini - Phone: +91 9941125984; E-mail: viji26priya@gmail.com; Corresponding author; Communicated by T. Lakshmi - lakshmi@saveetha.com

Received June 17, 2021; Revised September 27, 2021; Accepted September 27, 2021, Published December 31, 2021

DOI: 10.6026/973206300171105

Declaration on Publication Ethics:

The author's state that they adhere with COPE guidelines on publishing ethics as described elsewhere at <https://publicationethics.org/>. The authors also undertake that they are not associated with any other third party (governmental or non-governmental agencies) linking with any form of unethical issues connecting to this publication. The authors also declare that they are not withholding any information that is misleading to the publisher in regard to this article.

Author responsibility:

The authors are responsible for the content of this article. The editorial and the publisher have taken reasonable steps to check the content of the article in accordance to publishing ethics with adequate peer reviews deposited at PUBLONS.

Declaration on official E-mail:

The corresponding author declares that official e-mail from their institution is not available for all authors

This article is part of a special issue on Dental Biology

Abstract:

The development of antimicrobial drug resistance has encouraged scientists to develop alternate methods to combat infectious pathogens associated with dental diseases. Therefore, it is of interest to predict interactions for catechin (a plant derived compound) with protein targets in the red complex pathogens using computer aided network tools. However, *in vitro* and *in vivo* studies are warranted to confirm the antimicrobial effect of catechin (gallicocatechin, epicatechin, epigallocatechin (EGC) and gallolyl catechins) on the dental pathogens.

Keywords: Catechin, red complex pathogens, multi-drug transporters, efflux pumps.

Background:

Drug resistance to pathogens has reached alarming numbers in recent times. A global burden of antibiotic resistant organisms in the community as well as hospital settings is seen due to indiscriminate use of antibiotics. Oral micro flora is a complex environment wherein there is interplay between host and pathogens causing infectious diseases. Several pathogens of the oral cavity such as *Enterococcus faecalis*, *Streptococcus mutans* etc. are known to contain proteins responsible for drug resistant phenotypes [1,2]. Selective pressure rendered by the antibiotic laden environment is mainly responsible for the emergence and resurgence of such resistant forms.

Traditional or folk medicines have been used since decades to treat mild infections and disorders. The advantages of traditional medicine include (a) phyto compounds are derived from plants;

(b) non-toxic compared to synthetic drugs, (c) metabolized by the biological system with antimicrobial activity against pathogens. The red complex pathogens are a group of microorganisms mostly associated with severe periodontal infections. These organisms are usually found along with other bacteria in the periodontal pockets and cause destruction of periodontal tissues in a synergistic way [3]. Mouthwashes incorporated with bioactive principles from plants can be used as a substitute so as to break the communication between these pathogens. Several plant-derived compounds have been tested against various pathogens [4]. The biological properties of green tea have catechin including gallicocatechin, epicatechin, epigallocatechin (EGC), gallolyl catechins etc. EGC is the most abundant form and represents about 50% of all known types [5]. Bai *et al.* (2016) [8] showed the antimicrobial effect of epigallocatechin (EGCG) on canine oral bacteria. EGCG was found to possess a promising

growth and biofilm inhibiting property [6]. Several protein components of bacteria interacting with bioactive and synthetic compounds using computer-aided tools have been known [7,8]. Therefore, it is of interest to find the molecular targets of catechin on red complex pathogens.

Materials and Methods:

The mechanism underlying the anti-microbial activity of catechin against red complex pathogens is investigated using available computer aided predictive tools.

Strains used in the study:

The strains of red complex pathogens such as *Porphyromonas gingivalis* ATCC 33277, *Treponema denticola* ATCC 35405 and *Tannerella forsythia* ATCC 43037 available in the STITCH database were selected for the analysis (Figure 1, Table 1).

Analyzing protein interaction network:

The interactions between catechin and the protein repertoire of *Streptococcus mutans* UA159 and *Enterococcus faecalis* V583 was

used for predicting the functional class and virulent nature of the proteins [9]. The FASTA format of protein sequences was retrieved from the National Centre for Biotechnology Information (NCBI).

Prediction of functional class for interacting proteins:

VICMPred classifies the microbial proteins into four major classes such as (1) virulence factors; (2) information and storage processing; (3) cellular process and (4) metabolism. The virulence factors are identified using a support vector machine (SVM) algorithm, which classifies proteins based on their amino acid composition and sequence pattern [10].

Prediction of virulence properties of interacting protein:

VirulentPred is a yet another SVM based method, used for automated prediction of virulent proteins based on the sequences [11]. The scores with positive predicted values are more often categorized as virulent protein and those with negative predicted values are categorized as avirulent proteins.

Table 1: List of proteins of red complex pathogens interacting with catechin

Organism	Identifier	Proteins which interacts with Catechin	VICMPred Functional Class	VirulentPred	VirulentPred Score
<i>Porphyromonas gingivalis</i>	PGN_0081	Putative Na driven multidrug efflux pump	Metabolism	Avirulent	-1.020
	PGN_0677	Putative multi antimicrobial extrusion protein MatE	Metabolism	Avirulent	-1.016
	PGN_0490	Putative DNA-damage-inducible protein F	Metabolism	Avirulent	-1.005
	PGN_1207	Putative transport multidrug efflux protein	Metabolism	Avirulent	-1.024
	PGN_0006	Putative Sodium driven multidrug efflux pump	Metabolism	Avirulent	-0.981
<i>Tannerella forsythia</i>	BFO_2848	Multidrug transporter MatE	Metabolism	Avirulent	-1.119
	BFO_3258	Multidrug transporter MatE	Cellular Process	Avirulent	-1.015
	BFO_0631	Putative modification methylase HaeIII	Metabolism	Avirulent	-1.041
	BFO_2560	Hypothetical protein Tanf_11610	Metabolism	Avirulent	-1.023
	BFO_0443	GntR family transcriptional regulator	Cellular Process	Avirulent	-1.020
	BFO_0228	Multidrug transporter MatE	Virulence factor	Avirulent	-1.085
	BFO_0317	Multidrug transporter MatE	Metabolism	Avirulent	-1.032

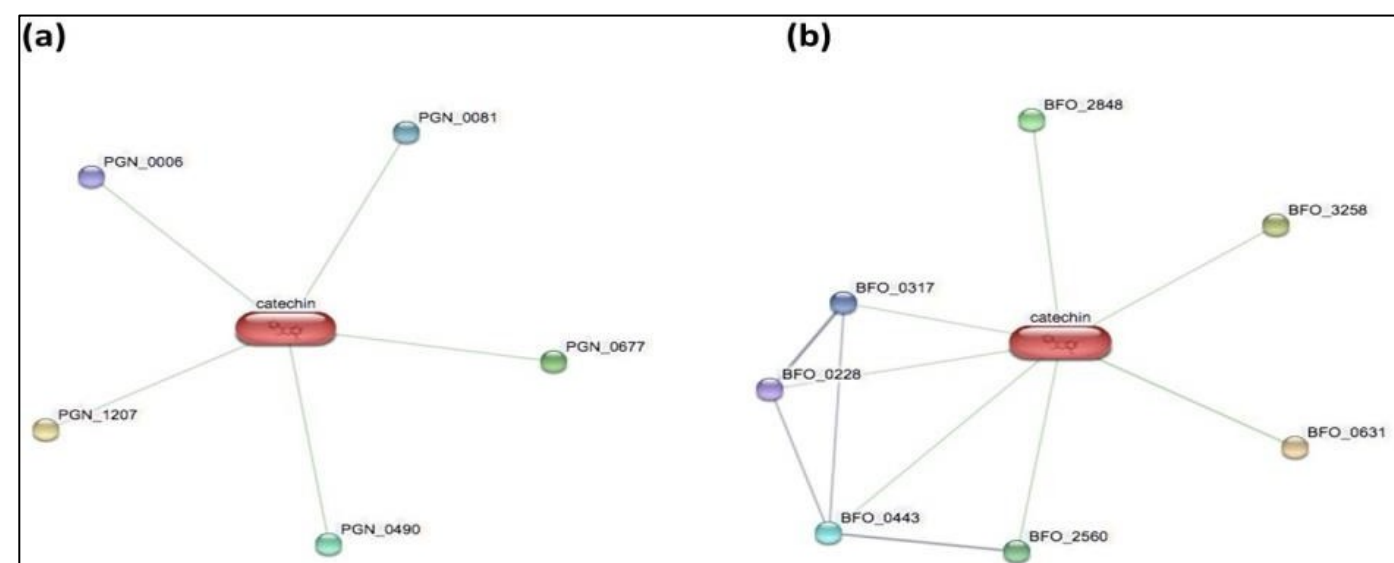


Figure 1: Interaction of catechin with the protein repertoire of red complex pathogens such as (a) *Porphyromonas gingivalis* and (b) *Tannerella forsythia*

Results and Discussion:

Catechin is a major component of green tea known for its antioxidant, anti-inflammatory, anti-proliferative activities. Bai *et al.* has shown the antimicrobial activity of catechin against a canine dental pathogen [8]. It is of interest to identify the possible molecular mechanisms underlying the antimicrobial effect of catechin using computational tools. The phytocompound catechin was found to target multiple proteins of *Porphyromonas gingivalis* and *Tannerella forsythia*. Data shows that catechin did not have interaction with the protein pool of *Treponema denticola*. However, the multidrug transporter MatE of *T. forsythia* was identified as a virulence factor using VICMPred tool and found to be avirulent by VirulentPred analysis. The MatE protein belongs to the multidrug efflux family protein. It is a transmembrane protein, which enables active transport of solutes by a mechanism. EGCG, a form of catechin has inhibitory activity on infections caused the multi-drug resistant *Staphylococcus aureus* [12, 13]. Falcinelli *et al.* showed the bactericidal effect of EGCG against both the attenuated *B. anthracis* and the virulent encapsulated *B. anthracis* Ames strain [14]. Maria *et al.* showed significant antimicrobial activity of EGCG against both gram negative (*Escherichia coli*) and gram positive (*Staphylococcus aureus*) [15]. The synergistic activity of catechin hydrate in combination with different antibiotics was tested against the clinical strains of *Staphylococcus aureus* is known. The results show that catechin hydrate produced significant antimicrobial activity in combination with clindamycin and erythromycin. These data shows the antibacterial effect of catechin on human pathogens.

Conclusion:

We document the predicted interaction targets for catechin (a plant derived compound) with protein targets in the red complex pathogens using computer aided network tools. However, *in vitro*

and *in vivo* studies are necessary to verify the antimicrobial effect of catechin (gallo catechin, epicatechin, epigallocatechin (EGC) and gallolyl catechins) on the dental pathogens.

Conflict of interest: The authors declare no conflict of interest.

References:

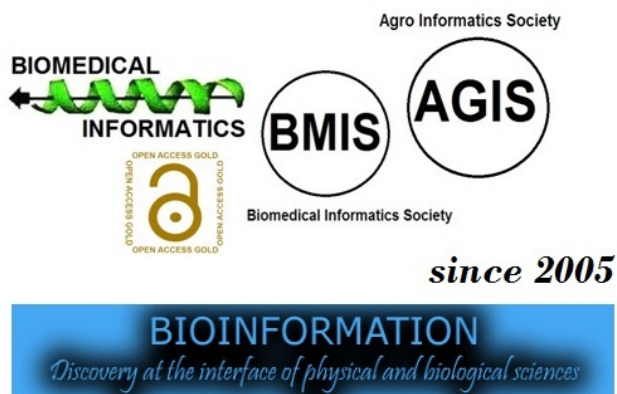
- [1] Vijayashree Priyadharsini J *et al.* *Heliyon*. 2018 **4**:e01051. [PMID: 30603692]
- [2] Priyadharsini JV *et al.* *Arch Oral Biol*. 2018 **94**:93. [PMID: 30015217]
- [3] Suzuki N *et al.* *Int J Dent*. 2013 **2013**:587279. [PMID: 23533413]
- [4] Pedrazzi V *et al.* *Scientific World Journal*. 2015 **2015**:712683. [PMID: 27239550]
- [5] Zou C *et al.* *Food Chem*. 2021 Nov 30 **363**:130322. [PMID: 34147900]
- [6] Ushanthika T *et al.* *Nat Prod Res* 2019:1. [PMID: 31311319]
- [7] Vijayashree PJ. *J Periodontol*. 2019 **90**:1441. [PMID: 31257588]
- [8] Bai L *et al.* *J Vet Med Sci*. 2016 **78**:1439. [PMID: 27246281]
- [9] Szklarczyk D *et al.* *Nucl Acid Res*. 2016 **44**:D380. [PMID: 26590256]
- [10] Saha S *et al.* *Gen Prot Bioinform*. 2006 **4**:42. [PMID: 16689701]
- [11] Garg A *et al.* *BMC Bioinform* 2008 **9**:62. [PMID: 18226234]
- [12] Stapleton PD *et al.* 2004 **23**:462. [PMID: 15120724]
- [13] Nance CL *et al.* *Aller. Clin. Immunol*. 2003 **112**:851. [PMID: 14610469]
- [14] Falcinelli SD *et al.* *FEMS Microbiol Lett*. 2017 **364**. [PMID: 28605495]
- [15] Mikłasińska M *et al.* *Molecules*. 2016 **21**:244. [PMID: 26907238]
- [16] Wu M *et al.* *Pathogens*. 2021 **10**:546. [PMID: 34062722]

Edited by P Kanguane

Citation: Cibikkarthik *et al.* Bioinformation 17(12): 1105-1108 (2021)

License statement: This is an Open Access article which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly credited. This is distributed under the terms of the Creative Commons Attribution License

Articles published in BIOINFORMATION are open for relevant post publication comments and criticisms, which will be published immediately linking to the original article for FREE of cost without open access charges. Comments should be concise, coherent and critical in less than 1000 words.



indexed in

