

# Identification of aspirin and diclofenac binding proteins in the red complex pathogens

Geethika Babu<sup>1</sup>, Veeraraghavan Vishnu Priya<sup>1,\*</sup>, Pothapur Keshav Krishna<sup>1</sup>, Rengasamy Gayathri<sup>1</sup> & Jayaseelan Vijayashree Priyadharsini<sup>2</sup>

<sup>1</sup>Department of Biochemistry, Saveetha Dental College, Saveetha Institute of Medical and Technical Sciences (SIMATS), Saveetha University, Chennai 600077, India; <sup>2</sup>Biomedical Research Unit and Laboratory Animal Centre-Dental Research Cell, Saveetha Dental College, Saveetha Institute of Medical and Technical Sciences (SIMATS), Saveetha University, Chennai 600077, India; \*Corresponding author - Veeraraghavan Vishnu Priya. Email id: vishnupriya@saveetha.com

Received December 29, 2020; Revised December 31, 2020; Accepted January 26, 2021, Published January 31, 2021

DOI: 10.6026/97320630017192

## Declaration on Publication Ethics:

The author's state that they adhere with COPE guidelines on publishing ethics as described elsewhere at <https://publicationethics.org/>. The authors also undertake that they are not associated with any other third party (governmental or non-governmental agencies) linking with any form of unethical issues connecting to this publication. The authors also declare that they are not withholding any information that is misleading to the publisher in regard to this article.

## Author responsibility:

The authors are responsible for the content of this article. The editorial and the publisher have taken reasonable steps to check the content of the article in accordance to publishing ethics with adequate peer reviews deposited at PUBLONS.

## Declaration on official E-mail:

The corresponding author declares that official e-mail from their institution is not available for all authors

## Abstract:

Red complex organisms are a group of organisms (*Porphyromonas gingivalis* ATCC 33277, *Treponema denticola* ATCC 35405, *Tannerella forsythia* ATCC 43037) that have been identified for the causation of periodontal diseases. Aspirin and diclofenac have been used as regular analgesics. Therefore, it is of interest to document the identification of aspirin and diclofenac binding proteins in the red complex pathogens using the STITCH v.5 pipeline. The virulence properties of these proteins were analyzed using VICMPred and VirulentPred software. Thus, we document 000 number of proteins having optimal binding features with the known analgesics.

## Background:

Periodontal infection is one of the most common dental infections that occur [1] second to dental caries. The first and initial process that is involved in the process of periodontitis is the colonisation of oral pathogens to form dental plaque [2]. There are various factors that are involved in the formation of biofilm such as the acidity of the oral environment and virulence of the bacteria and factors such as host immune response to inflammation plays a central role in

disease pathogenesis [3]. Periodontitis can be defined as an inflammatory disease of supporting tissues of teeth caused by specific microorganisms or groups of specific microorganisms, resulting in progressive destruction of the periodontal ligament and alveolar bone with periodontal pocket formation, gingival recession or both [4]. Recent advances have suggested that the varied microbial environmental present within the biofilms promotes accelerated genotypic and phenotypic diversity that provides a

form of cover that can safeguard the microbial colonies in the face of adverse conditions, such as those faced by pathogens in the host [2]. Periodontal diseases are basically bacterial infections associated with a complex microbial flora associated with that of the dental biofilm. This flora is composed predominantly of strictly anaerobic Gram-negative species that will in-turn produce a local and a systemic inflammatory response, ultimately leading to periodontal tissue destruction [5-7]. However, only few species such as *Aggregatibacter actinomycetemcomitans* (A.a) and *Porphyromonas gingivalis* have been classically grouped as periodontal pathogens [8,9]. Socransky et al. showed that periodontal diseases such as gingivitis and periodontitis are associated with a group of microbes rather than individual pathogens present at the periodontal sites. It comprises a consortium of three species namely, *Tannerella forsythia*, *P. gingivalis* and *Treponema denticola* which has been considered the most pathogenic microbial complex [5,10]. There have been various modalities of treatment of periodontitis, one of the treatment modalities is the use of therapeutic agents such as analgesics to obtain symptomatic relief, a few commonly used include aspirin and diclofenac for periodontitis [11,12].

#### Materials and Methods:

##### Study design:

The reaction as well as interaction of the compound with protein of bacteria was analyzed using STITCH v.5 pipeline<sup>13</sup> (Figure 1) and therefore the virulence properties of these interacting proteins were deduced and analysed by VICMPred<sup>14</sup> and VirulentPred softwares [15]. *Porphyromonas gingivalis* ATCC 33277, *Treponema denticola* ATCC 35405, *Tannerella forsythia* ATCC 43037 were the strains of red complex pathogens that were utilized in this study. These strains were included within the STITCH database, and therefore the query was user defined.

##### Prediction of protein-drug interactions:

STITCH database (Version 5; 2016) is an extensive platform for various predicted or known interactions between provides a comprehensive platform for known and predicted interactions between various compounds and proteins. The interactions between the compounds and the organism could vary from direct or physical and indirect or functional associations, which primarily arise from computational prediction and from interactions aggregated from various other (primary) databases (Figure 1). The repertoire of proteins, which interact with *P. gingivalis*, *T. denticola*, and *T. forsythia*, were further utilised for predicting virulence [13].

##### Virulence prediction:

VICMPred [14] and VirulentPred [15] pipelines were used for the identification of virulence factors targeted by aspirin and diclofenac among red complex pathogens. These tools employed support vector machine (SVM)-based five-fold cross-validation process to validate results. Virulence factors were screened on the idea of proteins using VirulentPred tool, which classified them into two groups' namely virulent and avirulent factors. VICMPred categorises proteins into four major classes, such as, proteins involved in cellular process, metabolism, information storage, and virulence. The general potent accuracies of VICMPred and VirulentPred servers were 70.75% and 86%, respectively. The FASTA formats of the actual proteins were retrieved from the NCBI database and were used as an input to run the algorithm (Figure 2) [15,16].

##### Prediction of subcellular localization of the virulent proteins:

The prediction of localisation of proteins at a sub cellular level helps in designing unique drug targets or for substantiating the role of an antimicrobial agent, which targets the virulent protein. Cell surface proteins are considered to be of great interest, as they will be used as vaccine targets. PSORTb V3.0 is an algorithm, which assigns a probable local site to a protein from an aminoalkanoic acid sequence that's provided [17]

##### Prediction of B-cell epitopes within the virulent proteins:

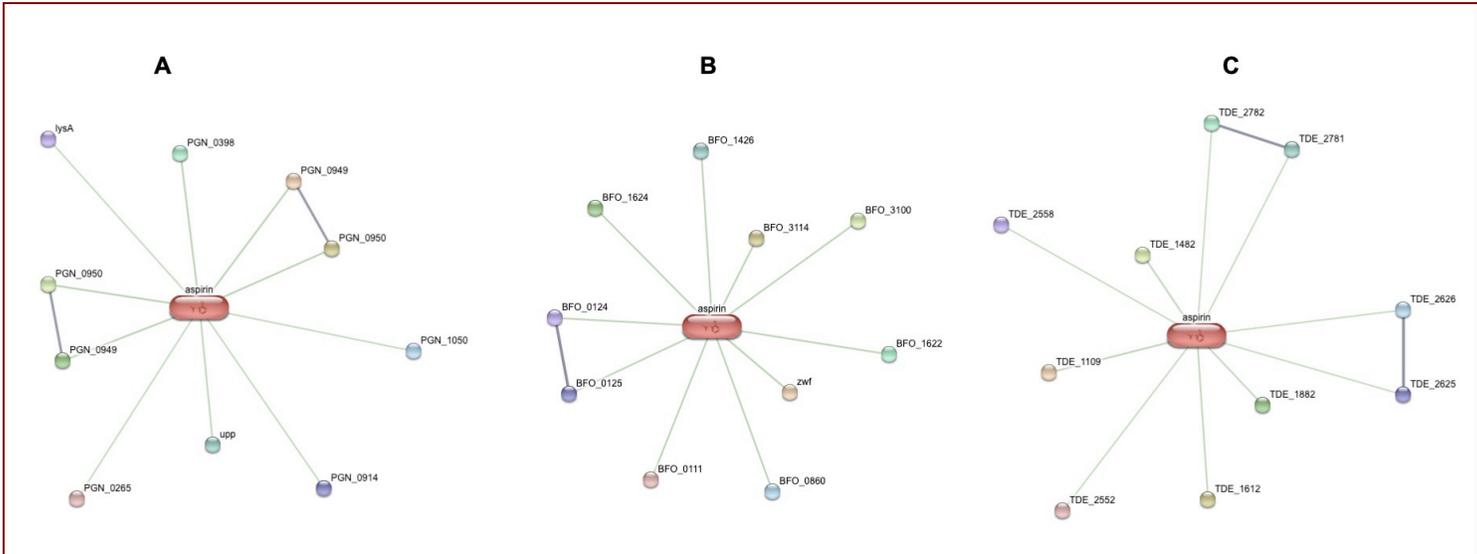
The BepiPred-2.0 server predicts B-cell epitopes from a protein sequence, employing a Random Forest algorithm on the idea of epitopes and non-epitope amino acids determined from crystal structures. The residues with scores above the edge (>0.5) are predicted to be a part of an epitope and colored in yellow on the graph (Figure 3) [18,19].

##### Results and Discussion:

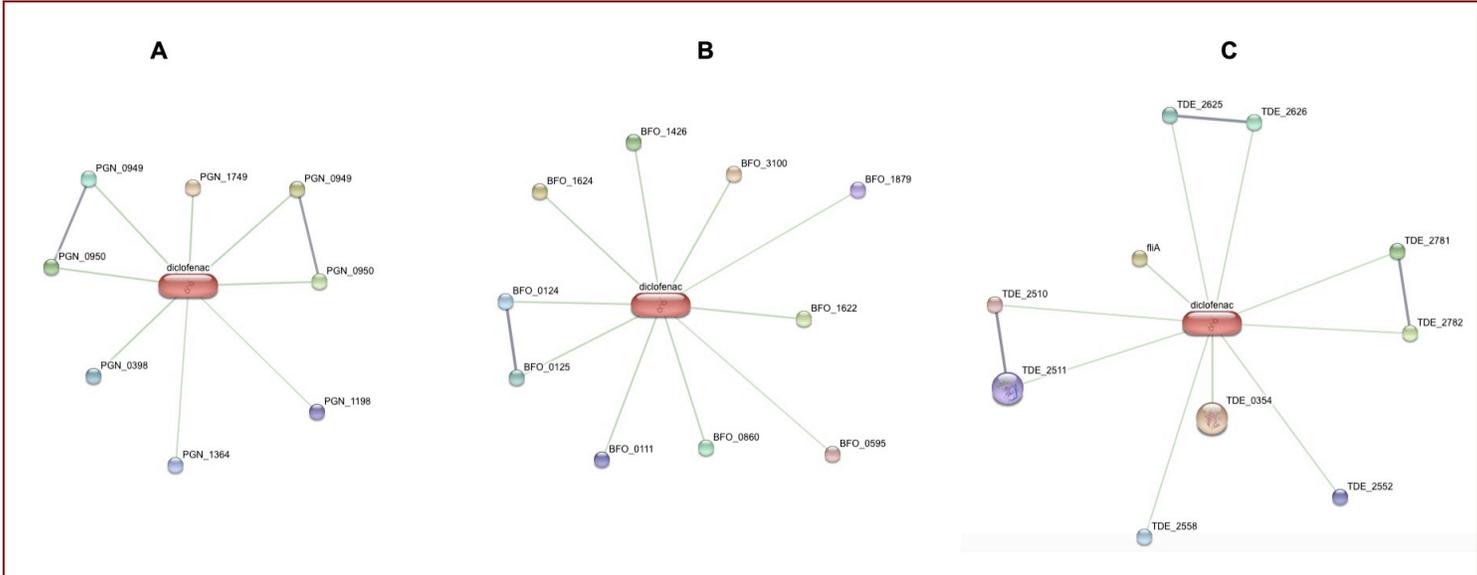
The STITCH pipeline was used to identify the protein interaction between red complex bacteria and the compounds, Aspirin and Diclofenac (Figure 1). Further each of the protein that was found to be interacting with the compounds was assessed for their virulence property using VirulentPred and VICMPred. The scores produced by the algorithms confirmed the nature of the proteins and grouped them into two classes, virulent and avirulent. Proteins interacting with aspirin were primarily related to metabolism processes, followed by cellular process. There were no proteins related to information storage or virulence factor that were identified that interacted with aspirin. The scores from VirulentPred marked carboxynorspermidine decarboxylase as virulent factors (Figure 1; Tables 1 and 2). Out of the proteins reacting with Diclofenac majority of them were related to metabolism followed by cellular

process and virulence factor. The scores from VirulenPred implied peptidyl-prolyl cis-trans isomerase cyclophilin type as the virulent Protein (**Figure 1; Tables 1 and 2**). STITCH prediction for Aspirin revealed proteins mainly associated with metabolism and cellular processes. None for virulence factor and information storage were retrieved. Two compounds such as Pyridoxyl dependent family decarboxylase and ABC transporter ATP-binding protein/permease, associated with metabolism were found to be virulent based on the score obtained from VirulentPred (**Figure 2; Tables 1 and 2**). When looked at interaction with Diclofenac most of the proteins that were retrieved belonged cellular process. Based on the scores obtained from VirulentPred ABC transporter ATP binding protein/ permease, related to metabolism was found to be virulent (**Figure 2; Tables 1 and 2**). Proteins interacting with aspirin, majority belonged to metabolism, followed by cellular process and virulence factor as retrieved from STITCH prediction. A protein, serpin associated with metabolism and a protein ABC transporter ATP-binding protein, associated with metabolism were predicted to be associated with virulence as per the scores obtained from VirulentPred. On interaction with Diclofenac, most of the interacting proteins belonged to metabolism. As per the scores obtained from VirulentPred, no protein was found to be virulent with respect to Diclofenac. Aspirin has been used through the ages in periodontitis patient and has been attributed to its anti-inflammatory property [11]. Aspirin has been used as an adjunct to standard periodontal therapy and has yielded good improvements in periodontal health [20,21]. This is a first of its kind study, as the antibacterial property of Aspirin has never been reported in literature against periodontal pathogens. Diclofenac has also been used in the post periodontal treatment and is highly debated for its usage attributed to its analgesic property [22,23]. The use of Diclofenac as an antibacterial agent has been reported against bacteria such as *Salmonella typhimurium* [24], *Escherichia coli* [25], and *Listeria* and *Mycobacterium tuberculosis*. [26] However the same hasn't been tested against red complex pathogens. Thus this study is a first of its kind in the field. Faizuddin *et al.* [27] has reported improvement in periodontal health and lesser periodontal attachment loss in patients with long-term aspirin usage. The improvement of periodontal health is attributed to reduced bleeding on probing which could have been due to the antiplatelet activity. The lower level of attachment loss is not accounted for and could be due to the modification of the bacterial flora with the intake of aspirin, but the same has to be evaluated clinically. Kim *et al.* [28] reported reduction in the pocket depth on usage of aspirin

for about a week and attributed the same to a reduction in inflammation attributed to the aspirin, the antibacterial property could also play a role in the same. In a study conducted by Fraser *et al.* [29] it has been reported that when antibiotic and aspirin as used against minor respiratory infections in two separate groups they yield similar results. This is proof to the fact that Aspirin has antibacterial property. Wang *et al.* [30] has reported anti bacterial property of aspirin against *H.pylori* and has reported that this could be due to the acidic nature of aspirin but that could not solely be the reason. They also reported that aspirin inhibits the growth of *H.pylori*. When in the diclofenac is considered, Milani *et al.* [31] have reported antibacterial property of diclofenac against *E.faecalis* which is one of the most common organisms involved in root canal treatment failure. The reason for the antimicrobial property of Diclofenac could be due to inhibition of bacterial DNA synthesis [32] or due to impairment of membrane activity [33,34] but the underlying mechanism is still unclear. In the present study these compounds have been screened for potential targets in red complex pathogens namely *P.gingivalis*, *T.denticola* and *T.forsythia*. Aspirin is found to be active against all three organisms whereas diclofenac is found to be inactive against *T.forsythia*. The protein ABC transporter ATP binding protein is found virulent in the case of aspirin and diclofenac with *T.denticola* and aspirin against *T.forsythia*. Matsson *et al.* [35] have reported novel drug delivery regarding this family of transporters, hence further studies can be carried to see if the same relation holds good in the case of periodontal pathogens. Vijayashree, 2019, has recently used the virtual screening methods to identify potential targets of non-steroidal anti-inflammatory drugs acetaminophen and ibuprofen against red complex pathogens [36]. Also, the authors analyzed certain phytochemicals using computational approach to identify targets in dental pathogens [37]. Hence, such in silico studies can reduce the time and cost required to assess the antimicrobial effect of bioactive compounds. The compounds validated using this procedure can be further screened using experimental procedures and clinical studies using animal and human volunteers. The present study is a unique who shows quintessential protein interactions of two commonly used drugs, aspirin and diclofenac against red complex pathogens. There are a few limitation of the study such that the reactions may be purely physical and may not hold any functional significance, the same protein interaction may not hold good in an in vivo environment, certain host proteins may mimic these proteins and thus should be taken care while using targeted drug therapy.



**Figure 1:** Interaction network of Aspirin with protein repertoire of red complex pathogens (A) *Porphyromonas gingivalis* (B) *Tannerella forsythia* and (C) *Treponema denticola*.



**Figure 2:** Interaction network of Diclofenac with protein repertoire of red complex pathogens (A) *Porphyromonas gingivalis* (B) *Tannerella forsythia* and (C) *Treponema denticola*.

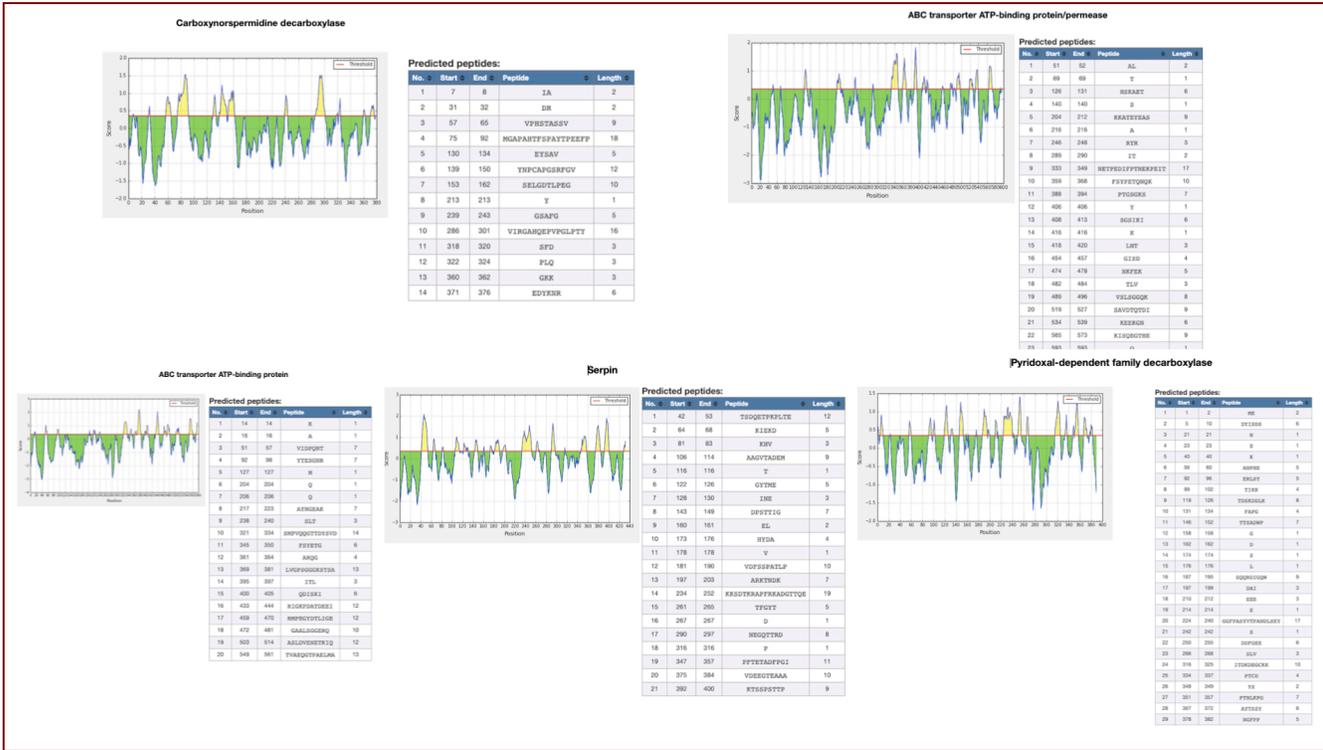


Figure 3: Epitopes identified in the virulent proteins of red complex pathogens interacting with Aspirin.

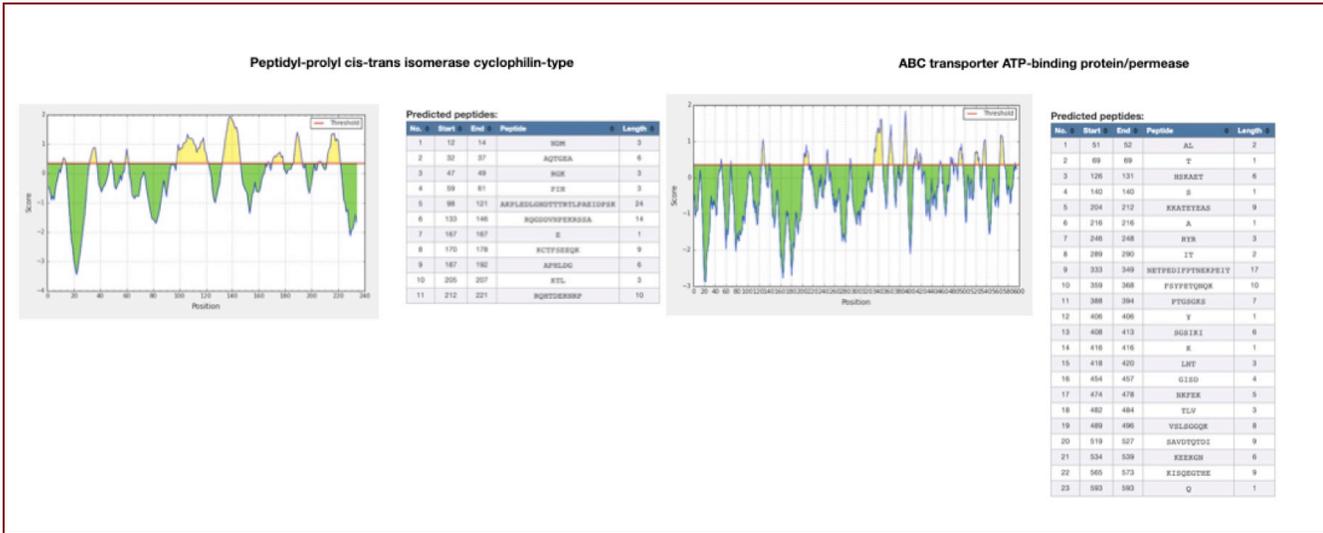


Figure 4: Epitopes identified in the virulent proteins of red complex pathogens interacting with diclofenac.

**Table 1:** Proteins of red complex pathogens interacting with acetylsalicylic acid (ASA) or Aspirin

Organism	Identifier	Proteins which interacts with luteolin	VICMPred Functional Class	VirulentPred	Virulent Pred Score
Porphyromonas gingivalis	PGN_0398	ABC transporter ATP-binding protein MsbA family	Cellular process	Avirulent	-0.605
	PGN_0949	ABC transporter ATP-binding protein	Metabolism	Avirulent	-1.234
	PGN_0950	ABC transporter ATP-binding protein	Metabolism	Avirulent	-1.220
	PGN_1050	Hypothetical protein	Cellular process	Avirulent	-1.472
	PGN_0914	Peptidase M24 family	Metabolism	Avirulent	-2.029
	Upp	Uracil phosphoribosyltransferase	Cellular process	Avirulent	-1.008
	PGN_0265	Carboxynorspermidine decarboxylase	Cellular process	Virulent	0.3589
	PGN_0949	ABC transporter ATP-binding protein	Metabolism	Avirulent	-1.234
	PGN_0950	ABC transporter ATP-binding protein	Metabolism	Avirulent	-1.220
	PGN_1272	Diaminopimelate decarboxylase	Metabolism	Avirulent	-1.128
Treponema denticola	TDE_2782	ABC transporter ATP-binding protein/permease	Metabolism	Virulent	0.5142
	TDE_2781	ABC transporter ATP-binding protein/permease	Cellular process	Avirulent	-1.024
	TDE_2626	ABC transporter ATP-binding protein/permease	Cellular process	Avirulent	-0.554
	TDE_2625	ABC transporter ATP-binding protein/permease	Cellular process	Avirulent	-0.791
	TDE_1882	Glycosyl hydrolase	Metabolism	Avirulent	-0.147
	TDE_1612	Phosphoribosyl transferase	Metabolism	Avirulent	-0.335
	TDE_2552	ABC transporter ATP-binding protein/permease	Cellular process	Avirulent	-0.207
	TDE_1109	Pyridoxal-dependent family decarboxylase	Metabolism	Virulent	0.3913
	TDE_2558	ABC transporter ATP-binding protein/permease	Metabolism	Avirulent	-0.999
	TDE_1482	Peptidase	Metabolism	Avirulent	-1.051
Tannerella forsythia	BFO_1426	ABC transporter ATP-binding protein	Cellular process	Avirulent	-1.191
	BFO_3114	Serpin	Metabolism	Virulent	0.1867
	BFO_3100	Putative lipid A export ATP-binding/permease protein MsbA	Virulence factor	Avirulent	-0.253
	BFO_1622	ABC transporter ATP-binding protein	Metabolism	Avirulent	-1.406
	BFO_0458	Glucose-6-phosphate dehydrogenase	Cellular process	Avirulent	-1.124
	BFO_0860	ABC transporter ATP-binding protein	Metabolism	Avirulent	-0.605
	BFO_0111	ABC transporter ATP-binding protein	Virulence factor	Avirulent	-0.970
	BFO_0125	ABC transporter ATP-binding protein	Metabolism	Avirulent	-0.170
	BFO_0124	ABC transporter ATP-binding protein	Metabolism	Avirulent	-1.295
BFO_1624	ABC transporter ATP-binding protein	Metabolism	Virulent	1.0005	

**Table 2:** Proteins of red complex pathogens interacting with diclofenac (DF)

Organism	Identifier	Proteins which interacts with DF	VICMPred Functional Class	VirulentPred	Virulent Pred Score
Porphyromonas gingivalis	PGN_1749	NADPH-quinone reductase	Cellular process	Avirulent	-1.302
	PGN_1917	ABC transporter ATP-binding protein	Metabolism	Avirulent	-1.234
	PGN_1916	ABC transporter ATP-binding protein	Metabolism	Avirulent	-1.220
	PGN_1198	Sodium-solute transporter	Cellular process	Avirulent	-1.187
	PGN_1364	Peptidyl-prolyl cis-trans isomerase cyclophilin-type	Virulence factor	Virulent	0.0256
	PGN_0398	ABC transporter ATP-binding protein MsbA family	Cellular process	Avirulent	-0.605
	PGN_0950	ABC transporter ATP-binding protein	Metabolism	Avirulent	-1.220
	PGN_0949	ABC transporter ATP-binding protein	Metabolism	Avirulent	-1.234
Treponema denticola	TDE_2625	ABC transporter ATP-binding protein/permease	Cellular process	Avirulent	-0.791
	TDE_2626	ABC transporter ATP-binding protein/permease	Cellular process	Avirulent	-0.554
	TDE_2781	ABC transporter ATP-binding protein/permease	Cellular process	Avirulent	-1.024
	TDE_2782	ABC transporter ATP-binding protein/permease	Metabolism	Virulent	0.5142
	TDE_2552	ABC transporter ATP-binding protein/permease	Cellular process	Avirulent	-0.207
	TDE_0354	General stress protein 14	Metabolism	Avirulent	-1.605
	TDE_2558	ABC transporter ATP-binding protein/permease	Metabolism	Avirulent	-0.999
	TDE_2511	ABC transporter ATP-binding protein/permease	Virulence factor	Avirulent	-0.72
	TDE_2510	ABC transporter ATP-binding protein/permease	Metabolism	Avirulent	-0.171
	fliA	RNA polymerase sigma factor WhiG	Cellular process	Avirulent	-1.162
Tannerella forsythia	BFO_1426	ABC transporter ATP-binding protein	Cellular process	Avirulent	-1.191
	BFO_3100	Putative lipid A export ATP-binding/permease protein MsbA	Virulence factor	Avirulent	-0.253
	BFO_1879	Cyclophilin type peptidyl-prolyl cis-trans isomerase	Cellular process	Avirulent	-1.886
	BFO_1622	ABC transporter ATP-binding protein	Metabolism	Avirulent	-1.364
	BFO_0595	Hypothetical protein	Metabolism	Avirulent	-1.277
	BFO_0860	ABC transporter ATP-binding protein	Metabolism	Avirulent	-0.605
	BFO_0111	ABC transporter ATP-binding protein	Virulence factor	Avirulent	-0.495
	BFO_0125	ABC transporter ATP-binding protein	Virulence factor	Avirulent	-0.970
	BFO_0124	ABC transporter ATP-binding protein	Metabolism	Avirulent	-0.170
	BFO_1624	ABC transporter ATP-binding protein	Metabolism	Avirulent	-1.295

**Table 3:** Subcellular localization of virulent proteins interacting with ASA and DF

Identifier	Protein	Subcellular location	Score
PGN_0265	Carboxynorspermidine decarboxylase	Cytoplasm	8.96
TDE_2782	ABC transporter ATP-binding protein/permease	Cytoplasmic membrane	10.00

TDE_1109	Pyridoxal-dependent family decarboxylase	Cytoplasm	9.97
BFO_3114	Serpin	Cytoplasm	8.96
BFO_1624	ABC transporter ATP-binding protein	Cytoplasmic membrane	10.00
PGN_1364	Peptidyl-prolyl cis-trans isomerase cyclophilin-type	Periplasm	9.76
TDE_2782	ABC transporter ATP-binding protein/permease	Cytoplasmic membrane	10.00

### Conclusion:

We document the identification of aspirin and diclofenac binding proteins in the red complex pathogens using the STITCH v.5 pipeline for further consideration.

### Conflicts of interest:

All authors declare that they have no potential conflict of interest for this work.

### References:

- [1] Oliver RC *et al.* *J Periodontol.* 1998 **69**:269.
- [2] Marsh PD. *BMC Oral health* 2006 **6**:pS14.
- [3] Yu OY *et al.* *Dent J.* 2017 **5**:21.
- [4] Newman MG *et al.* *Carranzas clinical Periodontology.* 10th ed. Elsevier health sciences; 2006.
- [5] Socransky SS *et al.* *J Clin Periodontol.* 1998 **25**:134.
- [6] Paster BJ *et al.* *Periodontol 2000.* 2006 **42**:80.
- [7] Page RC and Kornman KS. *Periodontol 2000* 1997 **14**:9.
- [8] Moore WE *et al.* *Infection and Immunity.* 1985 **48**:507.
- [9] Slots J and Ting M *Periodontol 2000.* 1999 **20**:82.
- [10] Holt SC *et al.* *Periodontol 2000.* 2005 **38**:72.
- [11] Schrodi J *et al.* *J Periodontol.* 2002 **73**:871.
- [12] Botelho MA *et al.* *Lat. Am. J. Pharm.* 2010 **29**:1371.
- [13] Szklarczyk D *et al.* *Nucleic acids research.* 2016 **44**:D380.
- [14] Saha S *et al.* *Genomics, proteomics & bioinformatics.* 2006 **4**:42.
- [15] Garg A, Gupta D. *BMC bioinformatics.* 2008 **9**:62.
- [16] <https://www.ncbi.nlm.nih.gov/protein>
- [17] Yu NK *et al.* *Bioinformatics.* 2010 **26**:1608.
- [18] Larsen JE *et al.* *Immunome Res.* 200 **24**:2.
- [19] Jespersen MC *et al.* *Nucleic Acids Res.* 2017 **45**:W24.
- [20] El-Sharkawy, H *et al.* *J Periodontol,* **81**:1635.
- [21] Drouganis A, Hirsch R. *J Clin Periodontol.* 2001 **28**:38.
- [22] Agarwal S *et al.* *Ind J Dent Res.* 2010 **21**:408.
- [23] Tramèr M *et al.* *Minerva stomatologica.* 2001 **50**:309.
- [24] Annadurai S *et al.* *Ind J Exp Biology.* 1998 **36**:86.
- [25] Mazumdar K *et al.* 2006 **20**:613. [PMID: 17091768].
- [26] Mazumdar K *et al.* *European journal of clinical microbiology & infectious diseases.* 2009 **28**:881.
- [27] Faizuddin M *et al.* *Australian dental journal.* 2012 **57**:45.
- [28] Kim DM *et al.* *J Periodontol* 2007 **78**:1620.
- [29] Fraser PK *et al.* *Lancet.* 1962 **614**.
- [30] Wang WH *et al.* *Gut* 2003 **52**:490. [PMID: 12631656]
- [31] Wang WH *et al.* *Gut* 2003 **52**:490. [PMID: 12631656]
- [32] Dastidar SG *et al.* *Int J Antimicrob Agents.* 2000 **14**:249.
- [33] Dutta NK *et al.* *Int J Antimicrob Agents.* 2007 **30**:242.
- [34] Dutta NK *et al.* *Int J Antimicrob Agents.* 2007 **30**:336.
- [35] Matsson P *et al.* *Pharm Res* 2009 **26**:1816.
- [36] Vijayashree Priyadharsini J. *J Periodontol.* 2019 **90**:1441.
- [37] Ushanthika T *et al.* *Nat Prod Res.* 2019.

Edited by P Kanguane

Citation: Babu *et al.* *Bioinformation* 17(1): 192-199 (2021)

**License statement:** This is an Open Access article which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly credited. This is distributed under the terms of the Creative Commons Attribution License

Articles published in BIOINFORMATION are open for relevant post publication comments and criticisms, which will be published immediately linking to the original article for FREE of cost without open access charges. Comments should be concise, coherent and critical in less than 1000 words.