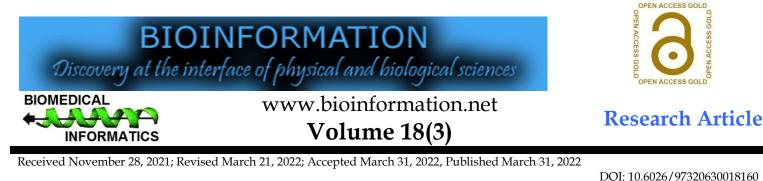
©Biomedical Informatics (2022)



Declaration on Publication Ethics:

The author's state that they adhere with COPE guidelines on publishing ethics as described elsewhere at https://publicationethics.org/. The authors also undertake that they are not associated with any other third party (governmental or non-governmental agencies) linking with any form of unethical issues connecting to this publication. The authors also declare that they are not withholding any information that is misleading to the publisher in regard to this article.

Declaration on official E-mail:

The corresponding author declares that lifetime official e-mail from their institution is not available for all authors

License statement:

This is an Open Access article which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly credited. This is distributed under the terms of the Creative Commons Attribution License

Comments from readers:

Articles published in BIOINFORMATION are open for relevant post publication comments and criticisms, which will be published immediately linking to the original article without open access charges. Comments should be concise, coherent and critical in less than 1000 words.

Edited by P Kangueane Citation: Harikrishnan *et al.* Bioinformation 18(3): 160-164 (2022)

Green synthesis and characterization of bisphosphonate conjugated gold nanoparticle with *Asparagus racemosus* root extract

Sruthi Harikrishnan¹, Navaneethan Ramasamy¹ & Rajeshkumar Shanmugam²

¹Department of Orthodontics and Dentofacial Orthopaedics, Saveetha dental college and Hospital, Saveetha Institute of Medical and Technical Sciences (SIMATS), Saveetha University Chennai-77, India; ²Department of Pharmacology, Saveetha Dental College and Hospital, Saveetha Institute of Medical and Technical Sciences (SIMATS) Saveetha University, Chennai - 77, India

Author contacts:

Sruthi Harikrishnan – E-mail:sruthident@gmail.com Navaneethan Ramasamy - E-mail:navaneethan@saveetha.com Rajeshkumar Shanmugam - E-mail:rajeshkumars.sdc@saveetha.com

Abstract:

Bisphosphonates improve orthodontic anchorage. More targeted action of this drug can be achieved through its conjugation with gold nanoparticles. *Asparagus racemosus* is a green edible medicinal plant used in Ayurvedic preparations to treat aging, vigor, immunity,

longevity, and skeletal issues. Therefore, it is of interest to report the green synthesized Bisphosphonate conjugated gold nanoparticles with *Asparagus racemosus* extract and to characterize them.

Keywords: Bisphosphonate; green synthesis; gold nanoparticles.

Background:

Nanotechnology aims to design, create and control matter in the dimensional range of 1-100nm [1]. The use of materials in these dimensions provides an opportunity to modify various properties such as solubility, diffusivity, blood circulation half-life, drug release characteristics, and immuno-genicity at the level of atomic or biomolecules [2, 3]. In general, synthesis of nanoparticles uses high radiation or concentrated reductants and stabilizing agents that are harmful both to the environment and to human health. Green synthesis of nanoparticles had shown to reduce the toxicity of the nanoparticles [4-7]. Bisphosphonates are increasingly being used to treat a wide range of skeletal disorders like osteoporosis. Bisphosphonates have been shown to enhance orthodontic anchorage in several studies [8-11]. Gold nanoparticles have several advantages, including the ability to accelerate osteoblast differentiation, inhibit adipose-derived stem cell differentiation, suppress osteoclast formation, and promote bone formation in bone tissue regeneration. When injected into the body, however, GNPs can cause toxicity. As a result, the surface of these particles must be modified to specifically target bone tissue [12-18]. Asparagus racemosus (A. racemosus, Shatavari) is a green edible medical plant used in Ayurvedic preparations to treat aging, vigor, immunity, longevity, and skeletal issues [19-21]. Bisphosphonate conjugated with gold nanoparticles has been shown to provide a more targeted action. Therefore, it is of interest to report the green synthesis and characterization of bisphosphonate conjugated gold nanoparticles with Asparagus racemosus root extract

Materials and Methods:

Green synthesis gold nanoparticles and conjugation of bisphosphonate:

Roots of Asparagus racemosus were dried in an oven at 30 °C and ground to a coarse powder. 1gm of available Asparagus racemosus stem powder was boiled at 100 degrees c with 100 ml of distilled water in a beaker. The extract was then filtered using a filter paper to obtain 75ml. To reduce the Au2+ ions, 30 mL of the extract was added to a reaction vessel containing 70 mL of chloroauric acid solution. The nanoparticles were then centrifuged at 1500 rpm for 10 minutes before being redispersed in 20 mL of distilled water. 2mg/ml of Zoledronic acid was added to one part of the sample gold nanoparticle extract and left to stir in a magnetic stirrer overnight.

Antimicrobial activity:

Biosynthesized nanoparticles are used in a wide range of biomedical applications. Membrane damage is one of the most common causes of nanoparticle antibacterial properties. Using the agar well diffusion method, the green synthesized bisphosphonate conjugated gold nanoparticles were tested against common oral pathogens such as *Candida albicans, Enterococcus fecalis, Staphylococcus aureus,* and *Lacto bacillus.* The test organisms (*S.* *mutans* and *Lactobacillus*) were grown in nutrient broth and kept on agar slants for the study. *Candida albicans* were grown on Rose Bengal agar, which is a yeast-specific medium. Using a sterile cotton swab, the freshly cultured strains were grown and uniformly spread over petri dishes containing MHS agar (Mueller Hinton 2 agar + 5% sheep blood). With the help of a steel borer, agar wells measuring 6.0 mm in diameter were punched into the culture plate containing the test microorganisms 35.A micropipette was used to fill the agar wells with 20µL of different concentrations of nanoparticles (50,100,150g/ml). As a positive control, 20µL of standard antibiotics (Ampicillin) were used. The diameter of the inhibition zone was measured in millimeters after a 24-hour incubation period at 37°C (mm). All of the tests were performed three times.

Cytotoxicity:

The eggs of brine shrimp are purchased to perform the cytotoxic assay on brine shrimp. The eggs are then kept at a temperature of 28°C. Artificial seawater and a 37°C light source are used to hatch eggs. This method was tested in 15 well plates (Figure 1)the newly hatched Nauplii are selected and transferred to each well using a Pasteur pipette. The Gold nanoparticles with and without Bisphosphonate conjugation were introduced into each of the wells of varying concentrations of 5,10,15,25 μ L is added to each well, and the volume is adjusted. After 24 hours, the brine shrimp are removed from the 15 well plates and counted with a magnifying glass. After a 24-hour incubation period, the percentage of dead shrimp in each well is calculated.

The number of motile nauplii was calculated to assess the cytotoxicity of the nanoparticles

Viability was calculated per well using the formula below

Viability (in %) =
$$\frac{\text{live brine shrimp after exposure}}{\text{live brine shrimp before exposure}}$$
 *100%

Result and Discussion:

The color change of the reaction mixtures from light yellow to yellow, dark-purple, and dark brown, respectively, could indicate the biosynthesis of Au nanoparticles in the current experiment. In the current experiment, the visual color change from yellow to dark purple was formed in 6 hours. The reduction confirmation of Au++ to Au0 is shown by the solution's color changing from light brown to dark brown. The brown color variation indicates an incomplete reduction of less concentration in the plant extract solution, whereas the formation of dark brown color at high plant extract concentrations revealed a complete reduction reaction. In the presence of incident photons, Au nanoparticles displayed the surface plasmon resonance (SPR) band as a result of the metal's conduction and free band electrons collectively oscillating. The

intensity of the SPR band is primarily determined by the nature of the nanoparticles used in the synthesis, as well as their composition Furthermore, UV-vis spectroscopy is a key tool for determining the nature of synthesized Au. The analysis was carried out every one hour to determine the changes. The analysis showed a consistent peak after 1 hr of preparation at 540 was constantly observed after 2 hours of the preparation of the sample (**Figure 1a and 1b**).

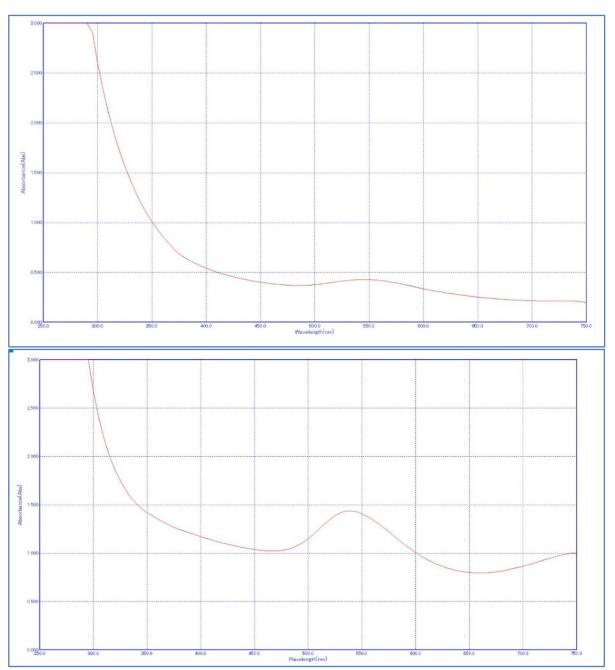


Figure 1: Shows the formation of the peak from 0 hr to 2 hr indicating the formation of the gold nanoparticles

Table 1: Antimicrobial activity of Bisphosphonate conjugated gold nanoparticles at various concentrations.

Nanoparticles	Organisms	& zone of ir	hibition for v	arying concent	rations of 1	nanoparticles	(ZOI) in mill	imeter (mm)							
	staphyloco	ccus aureus				Lactobacill	us				Candida al	bicans			
Bisphosphonate	25µg/ml	50µg/ml	100µg/ml	150µg/ml	control	25µg/ml	50µg/ml	100µg/ml	150µg/ml	control	25µg/ml	50µg/ml	100µg/ml	150µg/ml	control
conjugated gold	9	9	9	26	24	9	9	9	26	26	9	9	9	10	12
nanoparticles															

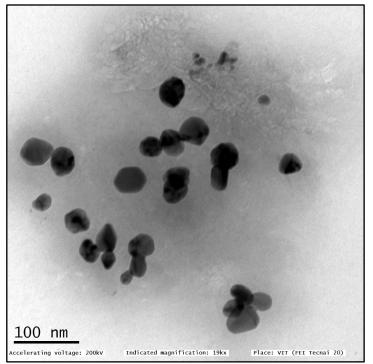


Figure 2: Transmission Electron Microscopic images of gold nanoparticles. AuNPs synthesized appear spherical, smooth, and measuring approximately 10–50 nm.

Table 2: Calculation of cytotoxicity at various concentrations of nanoparticles	s
---	---

Concentration (µg/mL)	No. of live nauplii (day 1)	No. of live nauplii (day 2)	% dead						
Control	10	10	0						
Gold nanoparticles without bisphosphonate									
5 μg/ml	10	10	0						
10 µg/ml	10	10	0						
15 µg/ml	10	10	0						
20 µg/ml	10	9	10%						
25 μg/ml	10	9	10%						
Gold nanoparticles with bisphosphonate									
5 µg/ml	10	10	0						
10 μg/ml	10	10	0						
15 µg/ml	10	10	0						
20 μg/ml	10	8	20%						
25 μg/ml	10	9	10%						

The TEM images and EDX spectra of biosynthesized Au nanoparticles showed that the particles are narrow in size and spherical in shape with a diameter in the range of 10–50 nm (Figure 2). However, some froth was noticed on the surface of these obtained nanoparticles, which could be attributed to the different types of phytochemicals present in the plant extract. FTIR analysis confirmed the presence of a huge amount of phytochemicals in the plant extract which can prevent the nanoparticles from agglomeration and helps in the production of stable nanoparticles. There was no other defined morphological difference observed in the preparation of Au nanoparticles. Antibacterial activity of biosynthesized bisphosphonate conjugated gold nanoparticle using *Asparagus racemosus* extract on *Staphylococcus aureus, Lactobacillus,* and *Candida albicans* using agar well diffusion method was

performed. This method is widely used to evaluate the antimicrobial activity of plants or microbial extracts. It is qualitative, easy to perform, and simple. The agar plate surface was inoculated by spreading a volume of the microbial inoculums over the entire agar surface. Then, a hole with a diameter of 6 mm was punched aseptically with a sterile cork borer or a tip, and a volume of 20 µL of the nanoparticle sample at desired concentration was introduced into the well. Then, agar plates were incubated under suitable conditions depending upon the test micro-organisms. The nanoparticle sample diffuses in the agar medium and inhibits the growth of the microbial strain tested. Four different concentrations of the nanoparticles were studied (25, 50,100,150µg/ml). The diameter of the zone of inhibition increased with an increase in the concentration of the nanoparticles, against bothS. aureus, Lacto bacillus, and Candida albicans. The diameter of the zone of inhibition against Enterococcus fecalis showed no change with the concentration of the nanoparticles. The zones of inhibition (in milli meters) of gold nanoparticles of varying concentrations, against S. aureus, Lactobacillus, and Candida albicans are represented in Table 1. Au nanoparticles have good antibacterial activity against Streptococcus mutans (150ug/ml - 26 mm zone of Inhibition), Lactobacillus (150ug/ml - 26 mm zone of Inhibition), and Candida albicans (150ug/ml - 10 mm zone of Inhibition). Cytotoxicity of the prepared nanoparticles was assessed using Brine Shrimp (Artemia salina) Lethality Assay. It has been demonstrated that the early developmental stages of Artemia salina are highly vulnerable to toxins. The lethality was found to be directly proportional to the concentration of the nanoparticles. Gold nanoparticles with and without bisphosphonate conjugation showed a mortality of 10% at 20 and 25 ug/ml (Table 2).

Conclusion:

We report the green synthesis and characterization of bisphosphonate conjugated gold nanoparticles with *asparagus racemosus* root extract for potential application in Dental Biology.

Funding: Nil

Informed consent statement: Not applicable.

Data availability statement: Data available on request.

Conflict of interest: There are no conflicts of interest.

References:

- [1] Zhang L et al. Clinical Pharmacology & Therapeutics 2008 83:761. [PMID: 17957183]
- [2] Brannon-Peppas et al. Advanced Drug Delivery Reviews 2004 56:1649. [PMID: 15350294]
- [3] Kawasaki ES *et al. Nanotechnology, Biology and Medicine.* 2005 1:101. [10.1016/j.nano.2005.03.002]
- [4] Anastas PT *et al.* Green Chemistry: Theory and Practice. Oxford University Press, USA; 2000. Page 135.
- [5] Kharissova OV *et al. Trends in Biotechnology.* 2013 31:240. [10.1016/j.tibtech.2013.01.003]
- [6] Narayanan KB et al. Advances in Colloid and Interface

ISSN 0973-2063 (online) 0973-8894 (print)

Bioinformation 18(3): 160-164 (2022)

©Biomedical Informatics (2022)

Science. 2011169:59. [10.1016/j.cis.2011.08.004]

- [7] Singh M et al. Research Journal of Nanoscience and Nanotechnology. 2011 1:1. [10.3923/rjnn.2011.1.11]
- [8] Nakas E et al. Bosnian Journal of Basic Medical Sciences. 2017 17:23 [10.17305/bjbms.2017.1715]
- [9] Fernández-González FJ *et al. Orthodontics and Craniofacial Research.* 2016 19:54. [10.1111/ocr.12115]
- [10] Seifi M et al. J Dent Res Dent Clin Dent Prospects. 2017 11:257. [29354254]
- [11] Putranto R *et al. Orthodontic Waves.* 2008 67:141. [10.1016/j.odw.2008.04.002]
- [12] Santhoshkumar J *et al. Biochemistry and Biophysics Reports.* 2017 11:46. [10.1016/j.bbrep.2017.06.004]
- **[13]** Shukla AK *et al. Green synthesis, Characterisation andappliacations of Nanoparticles*Elsevier; 2018[1]:548.

- [14] Dauthal Pet al. 3 Biotech. 20166 . [10.1007/s13205-016-0432-8]
- [15] Kus-Liśkiewicz M et al. Int J Mol Sci. 2021 20):10952. [34681612]
- [16] Fan L *et al. Nat Commun.* 2021 12:6371. [10.1038/s41467-021-26694-x]
- [17] Khanra K et al. Nano Biomedicine and Engineering. 2016 8:39-46[10.5101/nbe.v8i1.p39-46]
- [18] Pallela PNVK *et al. Applied Sciences.* 20191:421. [10.1007/s42452-019-0449-9]
- [19] Raut RW et al. Advanced Materials Letters. 2013.4:650. [10.5185/amlett.2012.11470]
- [20] Bopana N et al. J Ethnopharmacol. 2007 1:110. [17240097]
- [21] Lee D et al. Sci Rep. 2016 6:27336. [27251863]

