©Biomedical Informatics (2022)







www.bioinformation.net Volume 18(10)

Received September 2, 2022; Revised October 31, 2022; Accepted October 31, 2022, Published October 31, 2022

Declaration on Publication Ethics:

The author's state that they adhere with COPE guidelines on publishing ethics as described elsewhere at https://publicationethics.org/. The authors also undertake that they are not associated with any other third party (governmental or non-governmental agencies) linking with any form of unethical issues connecting to this publication. The authors also declare that they are not withholding any information that is misleading to the publisher in regard to this article.

Declaration on official E-mail:

The corresponding author declares that lifetime official e-mail from their institution is not available for all authors

License statement:

This is an Open Access article which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly credited. This is distributed under the terms of the Creative Commons Attribution License

Comments from readers:

Articles published in BIOINFORMATION are open for relevant post publication comments and criticisms, which will be published immediately linking to the original article without open access charges. Comments should be concise, coherent and critical in less than 1000 words.

Edited by P Kangueane Citation: Sonar *et al.* Bioinformation 18(10): 974-981 (2022)

Comparative docking analysis of tyrosine kinase inhibitors with HER2 and HER4 receptors

Priyanka Sonar¹, Karimunnisa Shaikh^{1*}, Sangeeta Ballav², Soumya Basu² & Sunil Harer³

¹Department of Pharmaceutics, Progressive Education Society's, Modern College of Pharmacy, Nigdi, Pune, M.S., India; ²Cancer and Translational Research Laboratory, Dr. D.Y. Patil Biotechnology and Bioinformatics Institute, Dr. D.Y. Patil Vidyapeeth, Tathawade, Pune, M.S., India; ³Department of Pharmaceutical Chemistry, Dattakala Shikshan Sanstha's, Dattakala College of Pharmacy, Pune, MS, India. *Corresponding author:

Author contacts:

Karimunnisa Shaikh - Email: karima78@rediffmail.com; Priyanka Sonar - E-mail: priyanka.harer@gmail.com

Abstract:

Tyrosine kinase receptors promote the growth and differentiation of normal breast and malignant human breast cancer cells, known as ERBB receptors. Various ERBB receptors are EGFR/ErbB1 and ErbB2/neu, which get over expressed in different solid tumors that activate upon binding of ligand to the extra cellular domain of these receptors. Of note, the epidermal growth factor receptor (EGFR) is a prime contributor to cancer through the involvement of four receptor tyrosine kinases (RTKs), namely, HER1, HER2, HER3, and HER4. Among them, HER2 and HER4 are majorly associated with breast cancer. Non-peptide quinazoline compounds homologous of the adenosine triphosphate (ATP) are competitively inhibited to RTKs to prevent cancer growth and metastasis. Various small drug molecule that targets the RTKs having the same scaffold, includes Lapatinib, Tivozanib, Erlotinib, Gefitinib, Crizotinib, and Ceritinib. The present study aims to

Research Article

DOI: 10.6026/97320630018974

investigate the comparative potential of structurally similar TKIs against HER2 and HER4 receptor receptors-silico molecular docking using FlexX software (LeadIT 2.3.2). Each docked complex's interaction profile was performed using BIOVIA Discovery Studio Visualizer 4.0. Molecular docking analysis was performed in order to get deeper insights into the interaction and binding pattern of the ligands with HER2 and HER4 receptors. The docking results revealed the Lapatinib compound acquired the relatively highest binding score of -32.36 kcal/mol and -35.76 kcal/mol with HER2 and HER4 proteins, respectively, concerning other compounds. Lapatinib is identified as a potential inhibitor for both the RTKs. Our study thus suggests the probable direction that could be further explored in inhibiting EGFR protein harboring breast cancer.

Keywords: HER 2, HER 4, molecular docking, tyrosine kinase receptors

Background:

The ERBB receptorsare tyrosine kinase family receptor that promotes growth and differentiation of both normal breast and malignant human breast cancer cells [1]. Epidermal growth factor receptor (EGFR/ErbB1), is one member of this family, over expressed in 20% to 80% of breast cancers [2, 3], and another member is HER2 (ErbB2/neu), is amplified and/or over expressed (i.e., HER2-positive) in 20% to 30% of breast cancers [4, 5]. EGFR and HER2 have emerged as promising targets for cancer therapy that drive tumor growth and progression. The EGFR family comprises four distinct membrane tyrosine kinase receptors; EGFR/ErbB-1, HER2/ErbB-2, HER3/ErbB-3 and HER4/ErbB-4, which are activated upon ligand binding to the extracellular domain of these receptors [6-7]. Formation of receptor homo- or hetero-dimers resulting in phosphorylation of tyrosine kinase residues and cross-phosphorylation, that triggers numerous signaling pathways such as phosphatidylinositol-3 kinase (PI3K), mitogen-activated protein kinase/extracellular signal-regulated kinases (MAPK/ERK1/2), signal transducer and activator of transcription (STAT), phospholipase C (PLCy), and/or the modulation of calcium channels [8], This sequence of events induces cellular responses which include proliferation, differentiation, and inhibition, of apoptosis, giving rise to diseases such as cancer [9]. The Tyrosine kinases are non-peptide aniline quinazoline compounds homologous of the adenosine triphosphate (ATP). This similarity allows them to compete for the ATP-binding domain of protein kinases preventing phosphorylation and subsequent activation of the signal transduction pathways, leading to apoptosis and decreasing cellular proliferation [10]. Under physiological conditions, the intrinsic activities of receptor tyrosine kinase inhibitors (RTKs) are strictly controlled [11]. Over expressed or increased activities of RTKs resulting from mutations, gene rearrangement or amplification have been correlated with tumor development and progression [12]. The epidermal growth factor receptor (EGFR), the first identified receptor of tyrosine kinases, is important for epithelial cell biology. It has been reported that EGFR is over expressed in various solid tumors such as gastrointestinal tract, non-small cell lung, breast, prostate, bladder, and ovarian carcinomas, head and neck cancers, and glioblastoma [13]. In the last decades, since the understanding of the key roles of RTKs in tumor development and progression, inhibition of RTK to prevent cancer growth and metastasis has become an attractive approach for the discovery of novel anticancer drugs [14]. The FDA has approved several small molecule receptor tyrosine kinase inhibitor drugs until September 2021, and additional inhibitors were approved by other regulatory agencies based on the various scaffold, that include Lapatinib, Tivozanib, Erlotinib, Gefitinib, Crizotinib, and Ceritinib [15]. Therefore, it is of interest to investigate the comparative potential of structurally similar TKIs against HER2 and HER4 receptors using molecular docking studies.

Materials and Methods:

Ligand and receptor preparation:

The SDF filesfor the drugs, Lapatinib (CID:208908), Erlotinib (CID: 176870), Gefitinib (CID:123631), Tivozanib (CID:9911830), Crizotinib (CID:11626560) and Ceritinib (CID:57379345) were downloaded from the PubChem database (https://pubchem.ncbi.nlm.nih.gov/) to serve as the docking ligands. These compounds were further prepared in Schrodinger Ligprepwizard. As Lapatinib is considered a potent inhibitor of HER2 and HER4 kinases, hence, it was used as a reference. The 3D X-ray crystal structure of two proteins; namely; HER2 kinase (ERBB2) (PDB: 3PP0) and HER4 kinase (ERBB4) (PDB: 3BBT) has resolutions of 2.25Å and 2.80 Å respectively were retrieved from Protein Data Bank (PDB)(https://www.rcsb.org/structure/2ZNN) [16, 17]. All of the bound ligands, water molecules, and other subunits were removed from both the proteins with the assignment of hydrogen atoms using the Protein Preparation Wizard (PPW) [18]. The positions of proteins and structure of ligands were minimized and optimized using the Optimized Potentials for Liquid Simulations-2005 (OPLS_2005) force field in Maestro [19]. Further, ionization states of the proteins were imparted by allotting amino acid chains using PROPKA program tool of PPW at pH 8.0.

Molecular docking study:

A molecular docking study was performedin order to explore the structural features of the ligands with the proteins using FlexX software (LeadIT 2.3.2) **[20]**. The active site of the protein was defined by selecting the amino acid residues around the resolutions 2.25Å and 2.80 Å of HER2 kinase (PDB: 3PP0) and HER4 kinase (PDB: 3BBT) respectively. The prepared ligands and proteins were fed into the docking protocol assigned with default parameters. The best conformation was determined by the binding affinity of ligands with the proteins. The interaction profile of each complex was performed by using BIOVIA Discovery Studio Visualizer 4.0.

Results and Discussion:

Taking into note, epidermal growth factor receptor (EGFR) is a prime contributor of cancer through the involvement of four receptor tyrosine kinases (RTKs), namely, HER1, HER2, HER3, and HER4.

©Biomedical Informatics (2022)



Figure 1: Molecular binding interaction of Tyrosine kinase inhibitors with HER2 receptor. Identified interaction profile of **(a)** 3PP0-Lapatinib **(b)** 3PP0-Tivozanib **(c)** 3PP0-Crizotinib **(d)** 3PP0-Gefitinib **(e)** 3PP0-Erlotinib. Representationas3D (left panel A) & 2D (right panel B).

ISSN 0973-2063 (online) 0973-8894 (print)

Bioinformation 18(10): 974-981 (2022)

©Biomedical Informatics (2022)



Figure 2: Molecular binding interactions of Tyrosine kinase inhibitors with HER4 receptor. Identified interaction profile of **(a)** 3BBT-Lapatinib **(b)** 3BBT-Tivozanib **(c)** 3BBT-Crizotinib **(d)** 3BBT-Gefitinib **(e)** 3BBT-Erlotinib **(f)** 3BBT-Ceritinib. Representation as 3D (left panel) & 2D (right panel).

Table 1: Different	docking interactions between	the ligands and HER2	receptor	D 11' ((0)
Ligand-Receptor	Docking score (kcal/mole)	Interacting residue	Interaction bond	Bond distance (A°)
3PP0-Lapatinib	-35.76	ASP808	Salt Bridge	1.90
		HOH	Halogen (F)	2.95
			Listerer (T)	2.68
		1 FIK/98 MET901	Flatogen (F)	2.02
		NIE1801 CED792	Conventional H-bond	1.88
		JER/03	Carbon H bond	2.04
		CI N700	Carbon H bond	2.52
		ASP863	Carbon H bond	2.51
		CED782	Halogon (E)	2.19
		JEN705	Pi sigma	2.01
		PHE864	Pi PiT shapod	5 30
		AI A751	Alkyl	4 15
		VAL734	Alkyl	4.15
		LYS753	Alkyl	3.92
		LEU726	Pi-Alkyl	519
		CYS805	Pi-Alkyl	4 39
		LEU726	Pi-Alkyl	5.28
		ALA751	Pi-Alkyl	3.73
		MET801	Pi-Alkyl	5.44
		LEU852	Pi-Alkyl	4 80
		LEU852	Pi-Alkyl	4.70
		VAL734	Pi-Alkyl	4.05
		ALA751	Pi-Alkyl	5.04
		LYS753	Pi-Alkvl	4.40
		LEU785	Pi-Alkyl	4.92
		LEU796	Pi-Alkyl	5.21
3PP0-Tivozanib	-22.70	ASP863	Conventional H-bond	1.96
		ASP863	Pi-sigma	2.37
		PHE864	Pi-PiT-shaped	5.41
		LEU726	Alkyl	4.79
		LEU796	Alkyl	4.70
		LEU785	Pi-Alkyl	4.95
		LEU796	Pi-Alkyl	4.44
		LEU726	Pi-Alkyl	4.78
		ALA751	Pi-Alkyl	4.72
		LEU852	Pi-Alkyl	4.06
		LEU726	Pi-Alkyl	3.67
		VAL734	Pi-Alkyl	3.97
		ALA751	Pi-Alkyl	5.21
		LEU852	Pi-Alkyl	5.01
3PP0-Crizotinib	-30.64	ASP808	Salt Bridge	1.78
		HO	Conventional H-bond	2.14
		MET801	Conventional H-bond	1.6
		GLN799	Conventional H-bond	1.83
		LEU800	Carbon H-bond	2.67
		GL Y804	Carbon H-bond	2.80
		ASP808	Carbon H-bond	2.73
		LEU726 LEU726	Carbon H-bond	2.92
		LEU720 MET901	Carbon H bond	2.55
		ASN850	Halogon (E)	2.55
		ASP863	Halogen (F)	2.83
		I FU726	Pi-sigma	2.05
		AL A751	Alkyl	4 39
		VAL734	Alkyl	4 27
		LEU726	Pi-Alkyl	5.07
		ALA751	Pi-Alkyl	4.10
		LEU800	Pi-Alkyl	5.33
		MET801	Pi-Alkyl	5.39
		LEU852	Pi-Alkyl	4.09
		VAL 724	Di Allavl	4.09

ISSN 0973-2063 (online) 0973-8894 (print)

Bioinformation 18(10): 974-981 (2022)

©Biomedical Informatics (2022)

3BBT-Gefitinib	-27.56	HOH	Conventional H-bond	2.59
		ASP808	Salt Bridge	1.73
		MET801	Conventional H-bond	2.02
		THR798	Carbon H-bond	3.04
		ASP808	Carbon H-bond	2.97
		GLN799	Carbon H-bond	2.59
		ASP863	Halogen (F)	3.61
		LEU726	Pi-sigma	2.52
		LYS753	Alkyl	4.46
		LEU796	Alkyl	5.13
		LEU726	Alkyl	3.99
		LEU726	Pi-Alkyl	5.32
		ALA751	Pi-Alkyl	3.54
		LEU852	Pi-Alkyl	4.14
		ALA751	Pi-Alkyl	5.24
		LEU852	Pi-Alkyl	4.58
		VAL734	Pi-Alkyl	4.91
		ALA751	Pi-Alkyl	5.33
		LYS753	Pi-Alkyl	4.71
3BBT-Erlotinib	-16.51	LYS753	Conventional H-bond	2.37
		ASP863	Carbon H-bond	2.98
		ASN850	Carbon H-bond	2.92
		ASP863	Carbon H-bond	2.37
		GLN799	Carbon H-bond	2.74
		MET801	Carbon H-bond	2.92
		MET801	Carbon H-bond	2.70
		ALA751	Carbon H-bond	2.40
		CYS805	Pi-sulfur	4.06
		VAL734	Pi-Alkyl	4.65
		VAL734	Pi-Alkyl	4.18
		ALA751	Pi-Alkyl	5.22
		LEU852	Pi-Alkyl	4.96

Table 2: Various docking interactions between the ligands and HER4 receptor

Ligand-Receptor	Docking score (kcal/mole)	Interacting residue	Interacting Bond		Bond distance (A°)
3BBT-Lapatinib	-32.36	GLU781	Carbon H-bond		2.42
-		GLN772	Carbon H-bond		4.30
		THR771	Carbon H-bond		2.58
		GLY777	Carbon H-bond		2.57
		CYS778	Conventional H-bond C	Conventional H-bond	2.95
		CYS778	Conventional H-bond		3.09
		THR771	Conventional H-bond		2.05
		MET774	Salt Bridge		2.17
		GLU781	Pi-Alkyl		1.60
		CYS778	Pi-Alkyl		4.32
		LEU669	Pi-Alkyl		5.03
		LEU825	Pi-Alkyl		4.53
		LEU825	Pi-Alkyl		4.73
		VAL707	Pi-Alkyl		5.38
		VAL756	Pi-Alkyl		5.50
		ALA724	Pi-Alkyl		3.41
		ALA724	Pi-Alkyl		2.70
		ALA724	Pi-Alkyl		5.29
		LEU699	Pi-Alkyl		5.03
		VAL707	Alkyl		4.53
		LEU769	Alkyl		4.66
		LYS726	Halogen (Cl,Br,I)		4.30
		LEU769	Pi-sigma		2.56
		LYS726	Pi-PiT-shaped		5.03
		PHE837	Pi-donor H-bond		5.13
		ASP836			2.66
3BBT-Tivozanib	-28.53	CYS778	Conventional H-bond		2.29
		ASP836	Conventional H-bond		1.94
		ASP836	Conventional H-bond		2.27
		ASP836	Conventional H-bond		2.05
		ASP836	Conventional H-bond		2.72
		MET774	Carbon H-bond		2.40
		PHE837	Pi-Pi T-shaped		4.20
		LEU669	Alkyl		5.32
		VAL707	Alkyi		4.28
		CY5778	Alkyl		4.37
		ME1747	Alkyl		3.69

ISSN 0973-2063 (online) 0973-8894 (print)

Bioinformation 18(10): 974-981 (2022)

©Biomedical Informatics (2022)

		LEU758	Alley	3.80
		EEC/38	Alkyl	5.80
		PHE837	Alkyl	4.26
		MET747	Pi-Alkyl	4.94
		I FU758	Pi-Alkyl	5.41
		LEO/30		5.41
		LEU669	P1-Alkyl	4.74
		ALA724	Pi-Alkyl	4.73
		I FU773	Pi-Alkyl	5 33
		LEOTIS		5.55
		LEU669	Pi-Alkyl	4.74
		CYS778	Pi-Alkyl	5.23
		VAL 707	D: Alleri	4.01
		VAL/0/	I I-MIKYI	4.21
		ALA724		4.36
		LEU825		5.47
3BBT_Crizotinih	24.87	CLU781	Salt Bridge	1.90
SDD1-CHZothinb	-24.07	GLU701	San bridge	1.90
		LYS726	Halogen (F)	2.04
		MET774	Conventional H-bond	1.30
		CI N772	Conventional H bond	1 72
		CLIN72		1.72
		CY5778	Conventional H-bond	1.94
		MET774	Carbon H-bond	1.83
		MET774	Carbon H bond	2.99
		IVIE 1774		2.99
		ILE725	Halogen (Cl,Br,I)	1.85
		ILE725	Halogen (F)	3.15
		LEU769	Halogen (F)	3 40
		NAL FOR		0.10
		VAL/0/	Аікуі	2.24
		LEU699	Pi-Alkyl	3.56
		I FU699	Pi-Alkyl	3.80
		LLCCOPP	D: 411 1	5.00
		ALA/24	PI-AIKyl	0.33
		LEU773	Pi-Alkyl	3.79
		LEU825	Pi-Alkyl	4 96
		NAL 707	D: A11-1	1.90 E 0E
		VAL/0/	Р1-АКУІ	5.05
		ALA724	Pi-Alkyl	5.38
		LYS726	Pi-Alkyl	4.99
2BBT Cofitinih	22.15	CLU781	Salt Bridge	1.02
3DD1-Gentinib	-22.13	GLU781	San bruge	1.93
		LYS/26	Conventional H-bond	3.02
		GLN772	Carbon H-bond	2.78
		MET774	Carbon H bond	2 70
		NIE1774		2.70
		ASP836	Halogen (F)	2.85
		GLY777	Pi-sigma	2.83
		AT A 724	Alley	4 32
		ILIO 24	11 IKy1	4.52
		LYS726	Alkyl	4.04
		LEU699	Alkyl	4.66
		AT A724	Pi-Alkyl	4 28
) (ETEZA	D: All 1	1.20 F 20
		ME1774	Р1-АКУІ	5.29
		LEU825	Pi-Alkyl	4.25
		LEU699	Pi-Alkyl	4 70
		LEUROE	D: A111	F 12
		LEU825	P1-AIKyI	5.13
		VAL707	Pi-Alkyl	4.51
		LYS726	Pi-Alkyl	5.09
3BBT-Erlotinih	-18.67	I VS726	Conventional H-bond	196
SDD1-Enotinio	-10.07	L 10720		1.90
		ME1774	Conventional H-bond	1.88
		CY778	Conventional H-bond	1.99
		ASP836	Conventional H-bond	2.19
		I EI 1772	Carbon H band	2.72
		LEU//S	Carbon H-bond	2.12
		GLN772	Carbon H-bond	2.77
		MET774	Carbon H-bond	2.37
		I VS726	Pi-sigma	2 70
		LIEFEO	A 11 - 1	1.00
		LUE/58	Аікуі	4.08
		LEU769	Alkyl	4.37
		VAL707	Pi-Alkyl	3.78
		VAL 707	D: All-1	2.00
		VAL/0/	PI-AIKyl	3.99
		LEU825	Pi-Alkyl	5.17
		VAL707	Pi-Alkyl	4.71
		AT A724	Di Allavi	1 10
		ALA/24		1.17
3BB1-Ceritinib	-20.65	ASP836	Attractive charge	5.12
		LYS726	Conventional H-bond	2.62
		CL V700	Halogon (Cl Br I)	1.64
		GLI/00		1.04
		GLY777	Carbon H-bond	1.81
		LEU769	Carbon H-bond	2.75
		I FI 1600	Halogon (Cl Br I)	2 / 2
		LEU079		2.42
		GLU781	Pi-Anion	3.48
		CYS778	Pi-sulfur	5.41
		AT A724	Albyl	1 37
		ALA/24		4.3/
		LEU825	Alkyl	4 32

©Biomedical Informatics (2022)

VAL7	'07 Alkyl	4.55
VAL7	'07 Pi-Alkyl	4.92
VAL7	'07	4.64
LEU8	25	5.14

Among them, HER2 and HER4 are majorly associated with breast cancer, and identification of their potential inhibitors is highly desired. Therefore, we performed molecular docking analysis in order to gain deeper insights into their interaction and binding pattern with the ligands. The docking results revealed that the reference compound, Lapatinib acquired the highest scores of -32.36 kcal/mol and -35.76 kcal/mol with HER2 and HER4 proteins respectively over all other compounds (Table 1). Our study identified Lapatinib as a potential inhibitor for both the RTKs. As shown in Figure 1 (a), Lapatinib is able to establish six Hydrogen bonds (H-bonds) within the HER2 protein active site; the first one between quinazoline nitrogen and MET801 (1.88 Å), the second one between guinazoline carbon and GLN799 (2.51Å), third one between fluorine halogen atom and THR798 (2.68 Å), fourth one between a carbofluoro of flourophenyl and SER783 (2.84 Å), fifth one between methoxy carbon and ASP863 (2.19 Å) and sixth one between sulfonyl hydrogen and ASP808 (2.52Å).

In molecular docking interaction with HER4 receptor, Lapatinib was seen to secure eight H-bonds; two between the fluorine halogen atom and THR771 (2.58 Å, 2.05 Å), the next two between the sulfonyl oxygen atom and CYS778 (2.95 Å, 3.09 Å) third one between quinazoline nitrogen and MET774 (2.17Å), one between the sulfonyl oxygen atom and GLY777 (2.57 Å), other one methoxy carbon and ASP836 (Å), next interaction between the oxygen atom of Furan and GLU781 (2.42Å), last one between guinazoline carbon and GLN772 (4.30 Å) (Figure 2). Comparatively, the other ligands did not exhibit good docking scores with the proteins while Erlotinib rendered the least affinity (Table 2). Besides, Ceritinib was found to be incompatible with the HER2 receptor and did not bind to it. Thus, the binding score order for HER2 protein was Lapatinib> Crizotinib> Gefitinib> Tivozanib> Erlotinib whereas for HER4 was Lapatinib > Tivozanib > Crizotinib > Gefitinib > Ceritinib > Erlotinib.

Conclusion:

Comparative docking study output of various tyrosine kinase inhibitors with Human epidermal growth factor receptor 2 (HER2) and HER2 suggest the promising binding response of Lapatinib over the other ligands. This study proposed that these drugs had represented the HER2 and HER4 receptors as targets of the epidermal growth factor receptor family. It is our aim to create increasing interest in utilizing these drugs in combination with chemotherapy and /or other HER2-directed agents in patients with central nervous system involvement, TKIs have shown to be effective in this setting for which treatment options have been previously limited and the prognosis remains poor. The aim of this study is to summarize the potential molecular docking interactions of currently approved TKIs for HER2+ breast cancer, and in nonsmall cell lung cancer supporting their use in key clinical trials, and in current clinical practice.

Funding:

This research work is supported by the Department of Science and Technology (DST) New Delhi, India. Grant Number [DST/WOSB/2018/1016/ETD/Priyanka (G), dated 17.10.2019].

Conflict of interest:

The authors report that there is no conflict of interest.

References:

- [1] Beiki O *et al. Breast Cancer Research.* 2012 **14**:R5 [PMID: 22225950]
- [2] http://globocan.iarc.fr
- [3] Perou CM *et al. Nature.* 2000 **406**:747 [PMID: 10963602]
- [4] Sotiriou C & Pusztai L. *New England Journal of Medicine*. 2009 **360**:790 [PMID: 19228622]
- [5] Slamon DJ et al. Science. 19892 44:707 [PMID: 2470152]
- [6] Dawood S et al. Journal of Clinical Oncology. 2010 28:92 [PMID: 19933921]
- [7] Brower V. Journal of National Cancer Institute. 2013 105:835 [PMID: 23733910]
- [8] Wang YN & Hung MC. Cell & Bioscience. 2012 2:13 [PMID: 22520625]
- [9] Yarden Y. European Journal of Cancer. 2001 37:S3-8 [PMID: 11597398]
- [10] Hossam M et al. Cancer Cell International. 2016 16:39 [PMID: 27231438]
- [11] Chong S & Bernards R. *Trends in Biochemical Sciences*.2014 **39**: 465 [PMID: 25239057]
- [12] Brunelleschi S et al. Current Pharmaceutical Design. 20028: 1959 [PMID: 12171522]
- [13] Ranson M. British Journal of Cancer. 2004 90:2250 [PMID: 15150574]
- [14] Zhang J et al. Nature Reviews Cancer. 2009 9:28 [PMID: 19104514]
- [15] Peng W et al. Trends in Pharmacological Sciences.2015 36:422[PMID: 25975227]
- [16] Aertgeerts K *et al. The Journal of biological chemistry.* 2011 286:18756 [PMID: 21454582]
- [17] Chen Q et al. Structure. 2008 16:460 [PMID: 18334220]
- [18] https://www.schrodinger.com/sciencearticles/protein-preparation-wizard.
- [19] Harder É et al. Journal of chemical theory and computation. 12:281 [PMID: 26584231]
- [20] Badry DB et al. Journal of Computer Aided Molecular Design.2003 17: 755 [PMID: 15072435]