



www.bioinformation.net
Volume 20(9)

Research Article

Received September 1, 2024; Revised September 30, 2024; Accepted September 30, 2024, Published September 30, 2024

DOI: 10.6026/9732063002001132

BIOINFORMATION 2022 Impact Factor (2023 release) is 1.9.

Declaration on Publication Ethics:

The author's state that they adhere with COPE guidelines on publishing ethics as described elsewhere at <https://publicationethics.org/>. The authors also undertake that they are not associated with any other third party (governmental or non-governmental agencies) linking with any form of unethical issues connecting to this publication. The authors also declare that they are not withholding any information that is misleading to the publisher in regard to this article.

Declaration on official E-mail:

The corresponding author declares that lifetime official e-mail from their institution is not available for all authors

License statement:

This is an Open Access article which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly credited. This is distributed under the terms of the Creative Commons Attribution License

Comments from readers:

Articles published in BIOINFORMATION are open for relevant post publication comments and criticisms, which will be published immediately linking to the original article without open access charges. Comments should be concise, coherent and critical in less than 1000 words.

Disclaimer:

The views and opinions expressed are those of the author(s) and do not reflect the views or opinions of Bioinformation and (or) its publisher Biomedical Informatics. Biomedical Informatics remains neutral and allows authors to specify their address and affiliation details including territory where required. Bioinformation provides a platform for scholarly communication of data and information to create knowledge in the Biological/Biomedical domain.

Edited by Vini Mehta

Citation: Parveen *et al.* Bioinformation 20(9): 1132-1135 (2024)

Impact of white tea extract - enriched calcium silicate cements on the flexural Strength and collagen degradation of dentin

Sufia Parveen¹, SJ Pragma², Alakesh Singha^{3,*}, Deeksha Goel⁴, Subasish Behera⁵ & Soumyaranjan Nanda⁶

¹Department of Conservative Dentistry and Endodontics, Buddha Institute of Dental Sciences and Hospital, Patna, Bihar, India;

²Department of Dentistry, ABVM Government Medical College, Rajnandgaon, Chhattisgarh, India; ³Private Practitioner, Department of Conservative Dentistry and Endodontics, Alaka's Odontocare Dental clinic & Endodontic Centre, Agartala, Tripura, India;

⁴Department of Conservative Dentistry and Endodontics, DJ College of Dental Science and Hospital, Niwari Road, Modinagar, U.P., India; ⁵Department of Conservative Dentistry and Endodontics, Government Dental College and Hospital (SCB), Manglabag, Cuttack, Odisha, India;

⁶Private Dental Practitioner, Conservative Dentistry and Endodontics, Cuttack, Odisha, India;

*Corresponding author

Affiliation URL:

<https://bidsh.org/>

<https://www.abvmgmcrajnandgaon.ac.in/>

<https://djdentalcollege.com/>

<https://scbdental.nic.in/>

Author contacts:

Sufia Parveen - E - mail: drsufia2k3@gmail.com

SJ Pragya - E - mail: drpragya191285@gmail.com

Alakesh Singh - E - mail: alakesh.singha2012@gmail.com

Deeksha Goel - E - mail: goeldeeksha858@gmail.com

Subasish Behera - E - mail: dr.subasish@gmail.com

Soumyaranjan Nanda - E - mail: nandasoumya245@gmail.com

Abstract:

The flexural strength of dentin and the rate at which dentin collagen degrades are two important variables that affect how long restorative dental materials last. The use of natural extracts to improve the qualities of dental materials has been studied recently. The antioxidant-rich white tea extract has the potential to be beneficial when mixed with calcium silicate cements. This *in vitro* research used forty removed human molar teeth. After the teeth were sectioned to obtain dentin specimens, the dentin was randomly divided into four groups (n=10) according to the type of treatment: Group A (control) consisted of the dentin that had not been treated; Group B treated the dentin using conventional calcium silicate cement; Group C treated the dentin using calcium silicate cement combined with 5% white tea extract; and Group D treated the dentin using calcium silicate cement combined with 10% white tea extract. A universal testing apparatus was used to measure flexural strength, and hydroxy-proline release analysis was used to measure collagen degradation during a 30-day period. ANOVA and post-hoc tests were used to examine the data, with a significance level of $p < 0.05$. Incorporating white tea extract into calcium silicate cements improves the flexural strength and lowers collagen degradation of dentin. The benefits are more noticeable with larger concentrations of extract.

Keywords: White tea extract, calcium silicate cement, dentin, flexural strength, collagen degradation, hydroxy-proline release, restorative dentistry

Background:

The overall strength and longevity of dental restorations are greatly influenced by the integrity of the dentin, especially its collagen matrix [1]. Over time, dental restorations may fail due to the mechanical qualities of dentin being compromised by collagen degradation, which is often made worse by restorative operations [2]. Because of its advantageous characteristics, including as their bioactivity, biocompatibility, and capacity to stimulate dentin regeneration, calcium silicate cements (CSCs) have found extensive use in restorative dentistry [3]. But one possible tactic to improve CSC function even more is to add natural extracts with antioxidant qualities to them [4]. Strong antioxidant and anti-inflammatory characteristics of white tea extract, which is high in polyphenols and flavonoids, may shield dentin collagen from enzymatic degradation [5]. Antioxidants have been shown in earlier research to suppress matrix metalloproteinases (MMPs) that break down collagen, protecting dentin's structural integrity [6]. Thus, the purpose of this work is to examine how the incorporation of white tea extract into CSCs affects dentin's flexural strength and collagen degradation.

Materials and methods:

Specimen Preparation: For this *in vitro* investigation, forty recently removed human third molars free of cavities and restorations were chosen. Before being used, the teeth were kept in a 0.9% saline solution at 4°C. Using a low-speed diamond saw

with water cooling, each tooth was sectioned transversely at the cements/enamel junction to create dentin slabs that were 2 mm thick. After that, the dentin slabs were polished to a uniform surface using 600-grit silicon carbide paper.

Experimental Groups: Based on the kind of treatment, the specimens were randomly assigned to four groups (n = 10). Group B had dentin treated with standard calcium silicate cement (ProRoot MTA, Dentsply), whereas Group A (Control) contained dentin that had not been treated. Dentin treated with calcium silicate cement and 5% white tea extract was used in Group C, whereas 10% white tea extract was added to dentin treated with the same cement in Group D. **Calcium Silicate Cement Preparation:** The manufacturer's instructions were followed in the preparation of the calcium silicate cement. White tea extract was added to the cement powder for Groups C and D at a weight percentage of 5% and 10%, respectively. In order to create the extract powder, white tea leaves were steeped in distilled water at 80°C for ten minutes. The tea leaves were then filtered and lyophilized. To get the cement formulations to the right consistency for applying on the dentin specimens, they were manually combined with distilled water.

Flexural Strength Testing: Using a universal testing machine (Instron, USA), a three-point bending test was performed on each dentin slab. The specimens were positioned atop two 10 mm-span supporting rods, and a load was applied at the slab's

center at a crosshead speed of 0.5 mm/min until the specimens failed.

Evaluation of Collagen Degradation: Over the course of 30 days, the amount of hydroxy-proline - a sign of collagen breakdown - that was released from the dentin specimens was measured. 5 mL of phosphate-buffered saline (PBS) were used to submerge each specimen, and it was then incubated at 37°C. Every 1, 7, 14, and 30 days, the PBS was collected and changed. Using a colorimetric test (Hydroxy-proline test Kit, Sigma-Aldrich) in accordance with the manufacturer's instructions, the amount of hydroxy-proline in the collected PBS was measured. A microplate reader was used to determine absorbance at 560 nm.

Statistical analysis:

IBM SPSS software, version 25.0, was used to analyze the data. For every group, the flexural strength and hydroxy-proline release mean and standard deviation were determined. To identify significant differences between the groups, a one-way ANOVA with post-hoc Tukey's test was used, with a threshold of statistical significance of $p < 0.05$.

Results:

Flexural strength:

The flexural strength values of the dentin specimens across the different groups are presented in **Table 1**. The incorporation of white tea extract into calcium silicate cement significantly increased the flexural strength of the dentin compared to the control group and the group treated with conventional calcium silicate cement. Group D (10% white tea extract) exhibited the highest flexural strength, followed by Group C (5% white tea extract), Group B (conventional calcium silicate cement), and Group A (control).

Table 1: Flexural strength of dentin specimens (MPa)

Group	Mean Flexural Strength (MPa)	Standard Deviation (MPa)
Group A (Control)	100.5	5.3
Group B (Conventional CSC)	105.8	4.7
Group C (5% White Tea Extract)	115.2	6.1
Group D (10% White Tea Extract)	120.4	5.9

Collagen degradation:

The hydroxy-proline release values, indicative of collagen degradation, for each group over the 30-day period are presented in **Table 2**. The groups treated with white tea extract (Groups C and D) showed significantly lower hydroxy-proline release compared to the control group and the group treated with conventional calcium silicate cement. The lowest collagen degradation was observed in Group D (10% white tea extract), followed by Group C (5% white tea extract), Group B (conventional calcium silicate cement), and Group A (control).

Table 2: Hydroxyproline release ($\mu\text{g/mL}$) Over 30 Days

Group	Day 1	Day 7	Day 14	Day 30
Group A (Control)	8.2	15.4	18.7	20.1
Group B (Conventional CSC)	7.6	14.2	17.3	18.9
Group C (5% White Tea Extract)	5.1	9.8	11.7	12.4
Group D (10% White Tea Extract)	4.7	8.6	10.2	10.8

Statistical analysis:

Statistical analysis revealed significant differences in both flexural strength and collagen degradation among the groups ($p < 0.05$). The flexural strength was significantly higher in Groups C and D compared to Groups A and B, with Group D showing the highest values. Similarly, the hydroxy-proline release was significantly lower in Groups C and D compared to Groups A and B, with Group D exhibiting the lowest values, indicating reduced collagen degradation. These results suggest that the incorporation of white tea extract into calcium silicate cement enhances the mechanical properties of dentin while simultaneously reducing collagen degradation.

Discussion:

The results of this investigation show that adding white tea extract to calcium silicate cement greatly improves dentin's flexural strength and slows down the deterioration of collagen. These results are in line with other studies that show antioxidant-rich natural extracts may enhance the mechanical qualities of dental materials [1]. The antioxidant activity of the polyphenols in the white tea extract may maintain the collagen matrix and shield it from enzymatic degradation, which explains the observed improvement in flexural strength in the extract-treated groups [2]. Given that the collagen matrix's structural integrity is essential for withstanding mechanical stress, this stabilization probably helps the dentin's mechanical integrity [3]. Furthermore, hydroxy-proline release, a measure of collagen degradation, significantly decreased, indicating that white tea extract efficiently inhibits matrix metallo-proteinases (MMPs), the enzymes that break down collagen in dentin [4]. Antioxidants have been shown in earlier research to be able to inhibit MMP activity, protecting the collagen network in dentin [5]. The effects were more noticeable at the higher concentration of white tea extract (10%), suggesting that the advantages seen were depending on the quantity of extract used. This dose-dependent impact is consistent with other research showing that dental materials with higher antioxidant concentrations might provide more protection [6]. The present findings underscore the potential of white tea extract as a beneficial additive in dental materials, particularly in enhancing the mechanical properties of calcium silicate cement. The significant improvement in flexural strength observed in the extract-treated groups may be attributed to the synergistic interaction between the bioactive compounds in white tea and the calcium silicate matrix. Previous studies have also highlighted the role of natural extracts in enhancing the mechanical properties of biomaterials by interacting with their inorganic components [7]. The polyphenolic compounds in white tea, known for their strong antioxidant activity, likely facilitate cross-linking within the collagen matrix; thereby reinforcing the structural integrity of dentin [8]. In addition to the mechanical benefits, the reduction in hydroxy-proline release suggests that white tea extract may play a crucial role in preserving the organic components of dentin. The inhibition of matrix metallo-proteinases (MMPs) by the extract could be one of the key mechanisms through which collagen degradation is slowed. This protective effect is

consistent with previous studies demonstrating the ability of antioxidants to modulate enzymatic activity, thereby preventing the breakdown of collagenous tissues [9].

The preservation of collagen is particularly important in dental applications, as it contributes to the overall durability and resilience of the dentin, which is critical for long-term dental health [10]. Moreover, the dose-dependent effect observed with varying concentrations of white tea extract indicates that optimizing the extract's concentration could be essential for maximizing its benefits in dental materials. Higher concentrations of white tea extract, such as the 10% used in this study, appear to provide more robust protection against collagen degradation and greater improvement in mechanical properties. This aligns with other research findings, which suggest that the concentration of bioactive compounds in natural extracts directly influences their efficacy in enhancing the properties of dental materials [11]. Future studies should explore the optimal concentration range to balance efficacy with material stability and handling properties [12]. The implications of these findings extend beyond the immediate application of white tea extract in calcium silicate cement. They suggest a broader potential for incorporating natural antioxidants into various dental materials to enhance their performance and longevity. This approach could lead to the development of more durable, biocompatible, and effective dental restoratives, particularly in areas where the preservation of dentin and prevention of collagen degradation are critical. The integration of such natural extracts could pave the way for more sustainable and health-promoting dental treatments, in line with the growing emphasis on biomimetic and regenerative dentistry [13]. Further research is needed to investigate the long-term effects of such materials in clinical settings, as well as their potential to interact synergistically with other dental treatments [14-16].

Conclusion:

Data shows that by strengthening dentin's flexural strength and slowing the deterioration of collagen, adding white tea extract to calcium silicate cement may increase the lifetime and durability of dental restorations. To investigate the long-term effects of white tea extract on dentin in clinical settings, further research is necessary.

References:

- [1] Peluso I & Serafini M, *Br J Pharmacol*. 2017 **174**:1195. [PMID: 27747873]
- [2] Cabrera C *et al*. *J Am Coll Nutr*. 2006 **25**:79. [PMID: 16582024]
- [3] Thompson JM *et al*. *J Endod*. 2012 **38**:62. [PMID: 22152622]
- [4] Sulkala M *et al*. *Arch Oral Biol*. 2007 **52**:121. [PMID: 17045563]
- [5] Stammers M *et al*. *J Biol Chem*. 2020 **295**:10562. [PMID: 32381510]
- [6] Rodríguez-Barragué J *et al*. *J Esthet Restor Dent*. 2021 **33**:702. [PMID: 33973710]
- [7] Yilmaz H *et al*. *J Prosthet Dent*. 2007 **98**:120. [PMID: 17692593]
- [8] Aguiar TR *et al*. *J Dent Res*. 2014 **93**:417. [PMID: 24574140]
- [9] AL, Ramesh S *et al*. *Cureus*. 2023 **15**:e48530. [PMID: 38074023]
- [10] Hardan L *et al*. *Cells*. 2022 **11**:2417. [PMID: 35954261]
- [11] Xiong Yet *et al*. *J Dent Sci*. 2022 **17**:1135. [PMID: 35784122]
- [12] Kharouf Net *et al*. *Bioengineering (Basel)*. 2020 **7**:72. [PMID: 32645860]
- [13] Gascón R *et al*. *Materials (Basel)*. 2023 **16**:2260. [PMID: 36984145]
- [14] Tjäderhane Let *et al*. *Dent Mater*. 2013 **29**:999. [PMID: 23953737]
- [15] Davari A *et al*. *J Dent (Shiraz)*. 2013 **14**:136. [PMID: 24724135]
- [16] Cortez TV *et al*. *Restor Dent Endod*. 2024 **49**:e9. [PMID: 38449495]